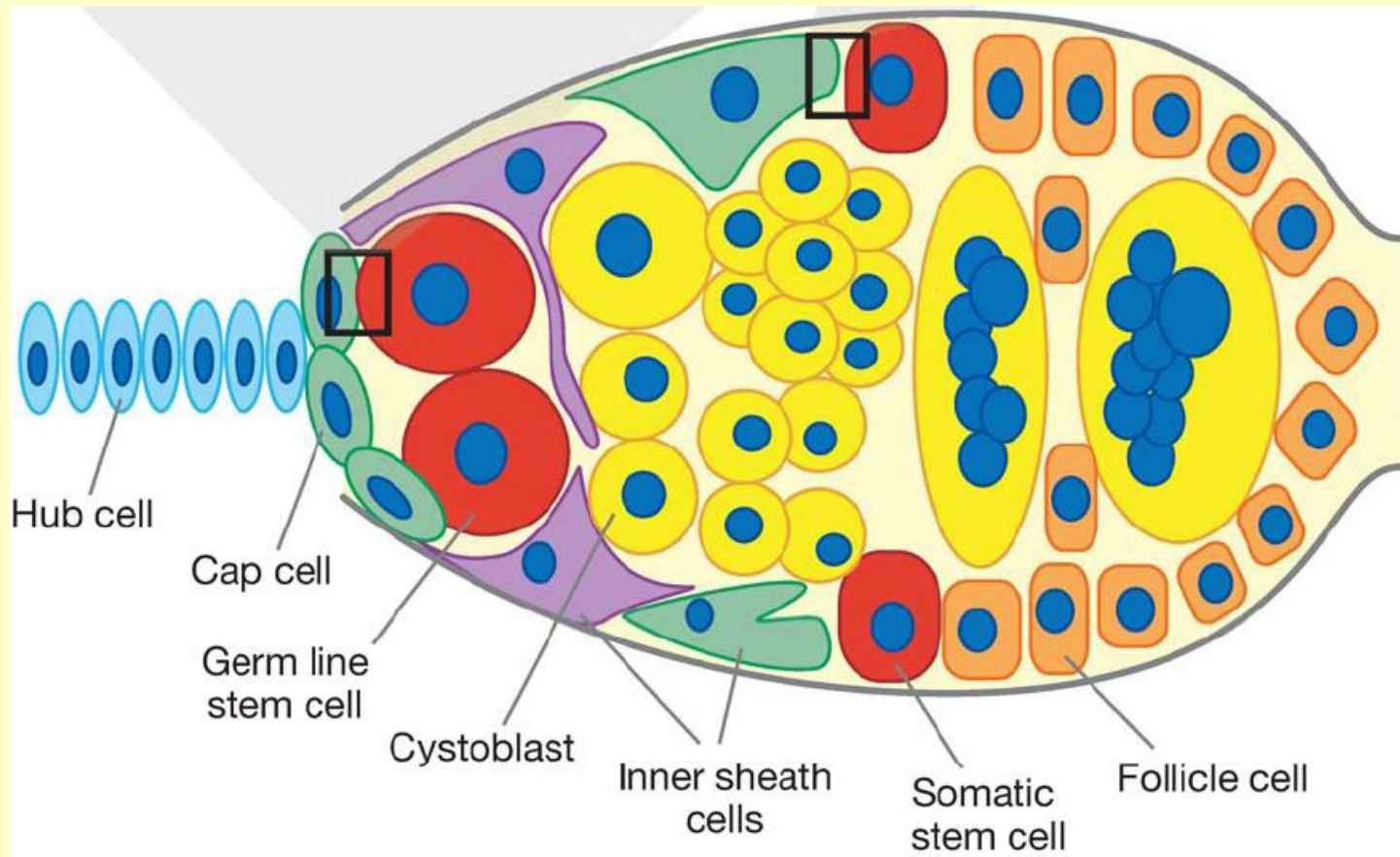


# Genomics & Medicine

<http://biochem118.stanford.edu/>

## Stem Cells

<http://biochem118.stanford.edu/Stem Cell.html>



Doug Brutlag, Professor Emeritus of  
Biochemistry & Medicine (by courtesy)  
Stanford University School of Medicine

# Causal Mutation Homework Assignment

Most of the SNP variations associated with diseases in genome-wide association studies do not cause the disease, but instead, these SNPs serve as genetic markers that are linked to genes which are involved in the disease. Ongoing research is attempting to sequence these genes in patients and in controls to find the actual variations in these genes that do in fact, cause the disease.

For this assignment I would like you to choose a simple Mendelian inherited disease other than those mentioned in class (Huntingtons, diabetes, Parkinsons, cystic fibrosis, sickle cell, etc.) and describe what is known about the genetic variations that cause that disease.

You may search [OMIM](#), [dbSNP](#), [dbVAR](#), [HGMD](#), [HGVS](#), [ClinVar](#), [SwissVar](#) and other database of genome variations that are associated with specific diseases to find an example of the kinds of mutations associated with the disease. Please describe how each of these variations cause the disease.

Is it by:

- 1) mutating the coding region of the protein
  - 2) altering the gene expression by affecting the promoter
  - 3) altering gene expression by affecting a transcription factor binding site
  - 4) altering gene expression indirectly by mutating a transcription factor itself
  - 5) altering copy number, hence changing gene expression levels
  - 6) altering other regulatory sites (miRNA targets)
  - 7) altering splice signals
- etc.

Often there will be several types of mutations that can cause the disease. Please comment on all types that are known for your chosen disease.

## Welcome to **Henry Stewart Talks Online Collections**

We have over 2,000 online seminars by leading world experts, covering biomedicine, life sciences, management, marketing, finance and many more topics.

### The Biomedical & Life Sciences Collection



### The Marketing & Management Collection




## Series: **Stem Cells**

Recent advances in understanding and utilizing

### TOPICS COVERED

- Embryonic stem cells in perspective
- The advent of direct reprogramming
- Embryonic stem cells: derivation and properties
- Pluripotency and disease modeling: insights into reprogramming mechanisms
- Stem cells and regeneration: The physiological function of mesenchymal stem cells
- Niche regulation of stem cell function: stem cells and tissue homeostasis
- RNA regulation and stem cells: microRNA regulation of pluripotency
- The aging of mitotic cells: regeneration and aging
- Stem cells and cancer: lineage tracing in normal stem cells and cancer
- Stem cells derived from amniotic fluid and placenta
- Cord blood stem cells
- Mesenchymal stem cells derived from bone marrow
- Hematopoietic stem cells
- Stem cells derived from peripheral blood
- Stem cells derived from fat
- Skeletal muscle stem cells
- Epithelial skin stem cells
- Stem cells and heart disease
- Islet cell therapy and pancreatic stem cells
- Cell therapy of liver disease: from hepatocytes to stem cells



# HumBio 157

## The Biology of Stem Cells

---

### **HUMBIO 157: The Biology of Stem Cells (DBIO 257)**

The role of stem cells in human development and potential for treating disease. Guest lectures by biologists, ethicists, and legal scholars. Prerequisites: HumBio 2A and 3A, or the equivalent in the BioCore in Biological Sciences.

**Terms:** Spr | **Units:** 3 | **UG Reqs:** WAY-SMA | **Grading:** Letter or Credit/No Credit

**Instructors:** Fuller, M. (PI) ; Nusse, R. (PI)

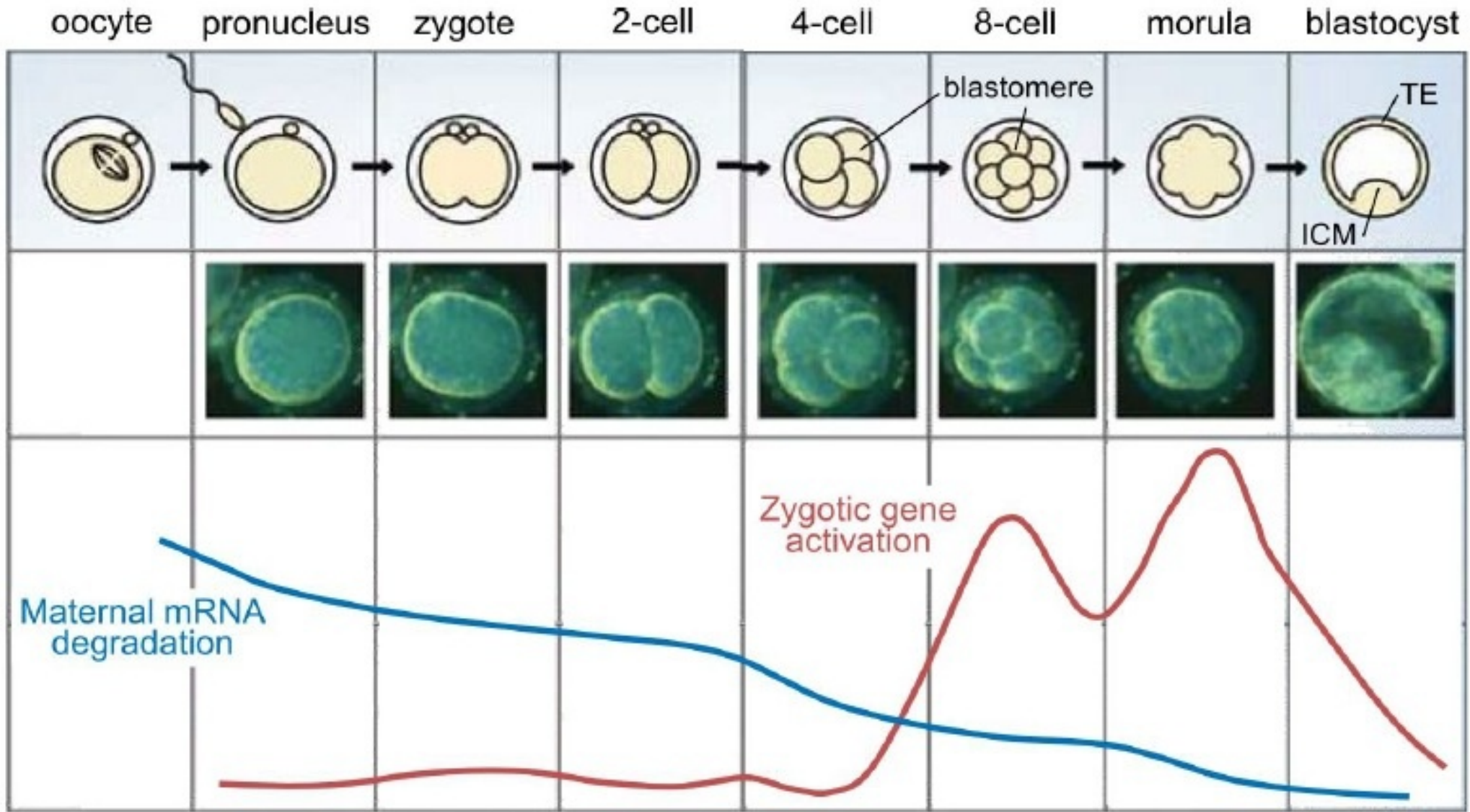
[Schedule for HUMBIO 157](#)

### **2014-2015 Spring**

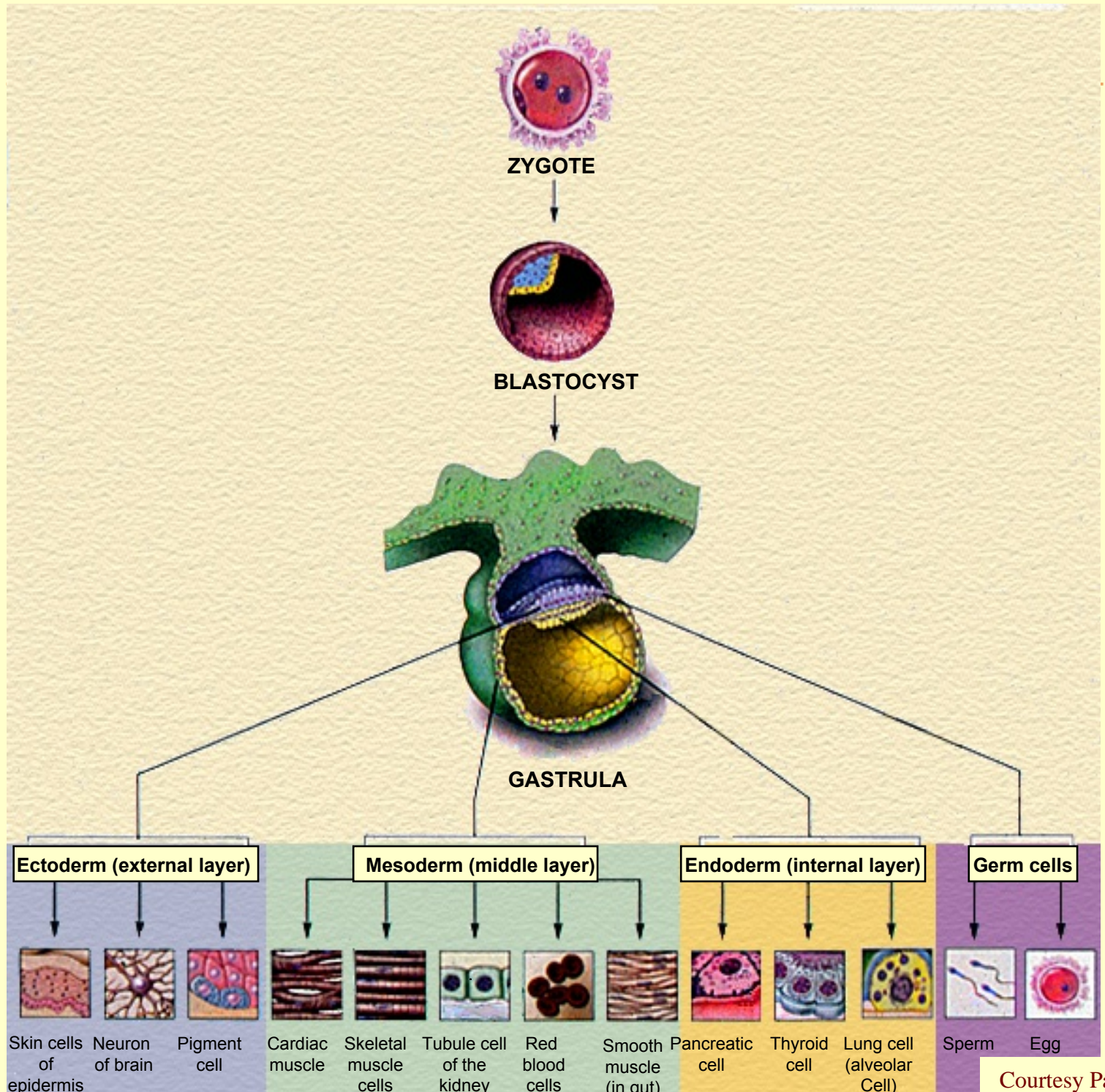
HUMBIO 157 | 3 units | UG Reqs: WAY-SMA | Class # 20511 | Section 01 | Grading: Letter or Credit/No Credit | LEC  
03/30/2015 - 06/03/2015 Tue, Thu 2:15 PM - 3:45 PM with Fuller, M. (PI); Nusse, R. (PI)

**Instructors:** Fuller, M. (PI); Nusse, R. (PI)

# Early Embryo Development

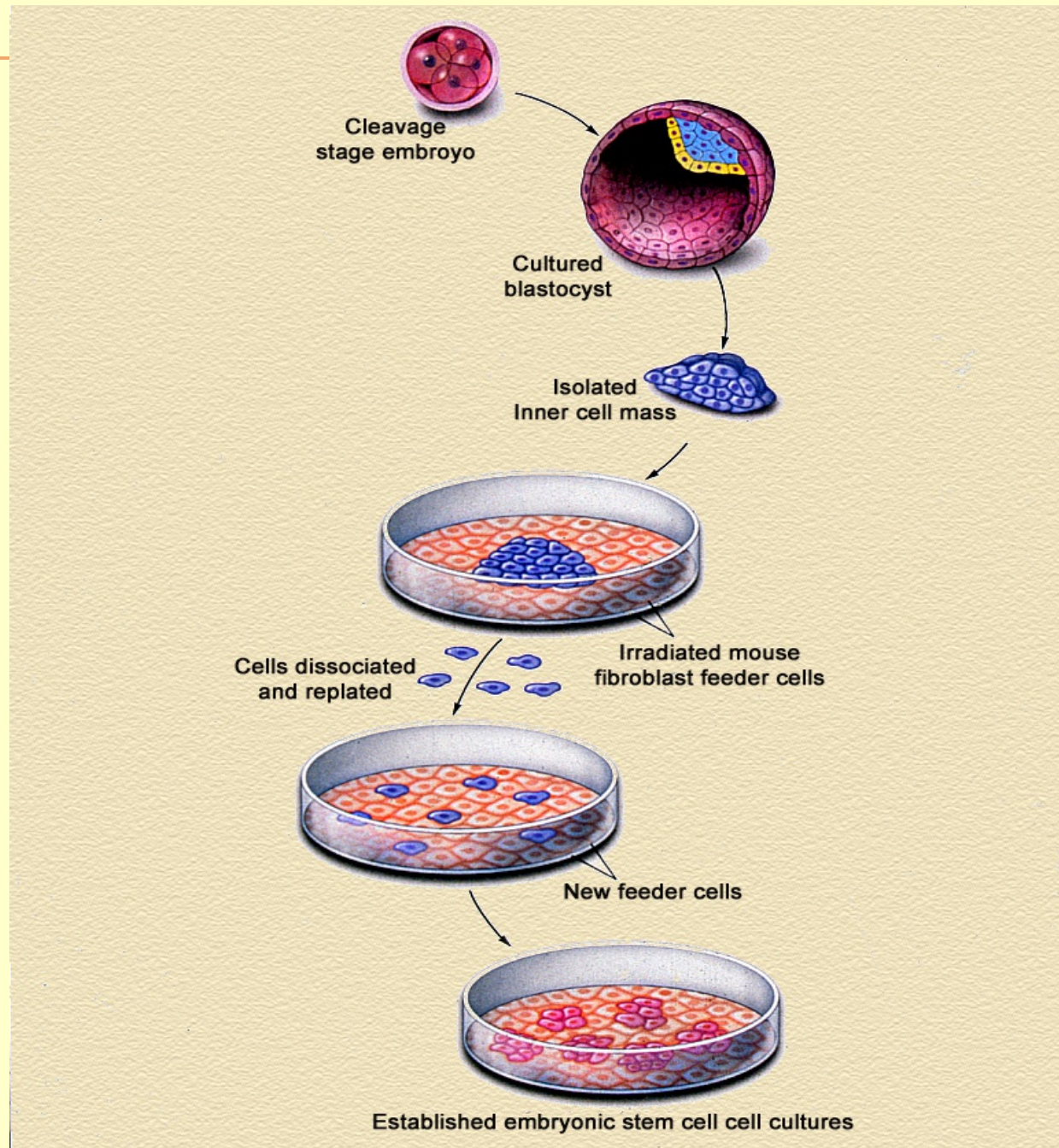


# Differentiation of Human Tissues



Courtesy Paul Berg

# Embryonic Stem Cell Cultures

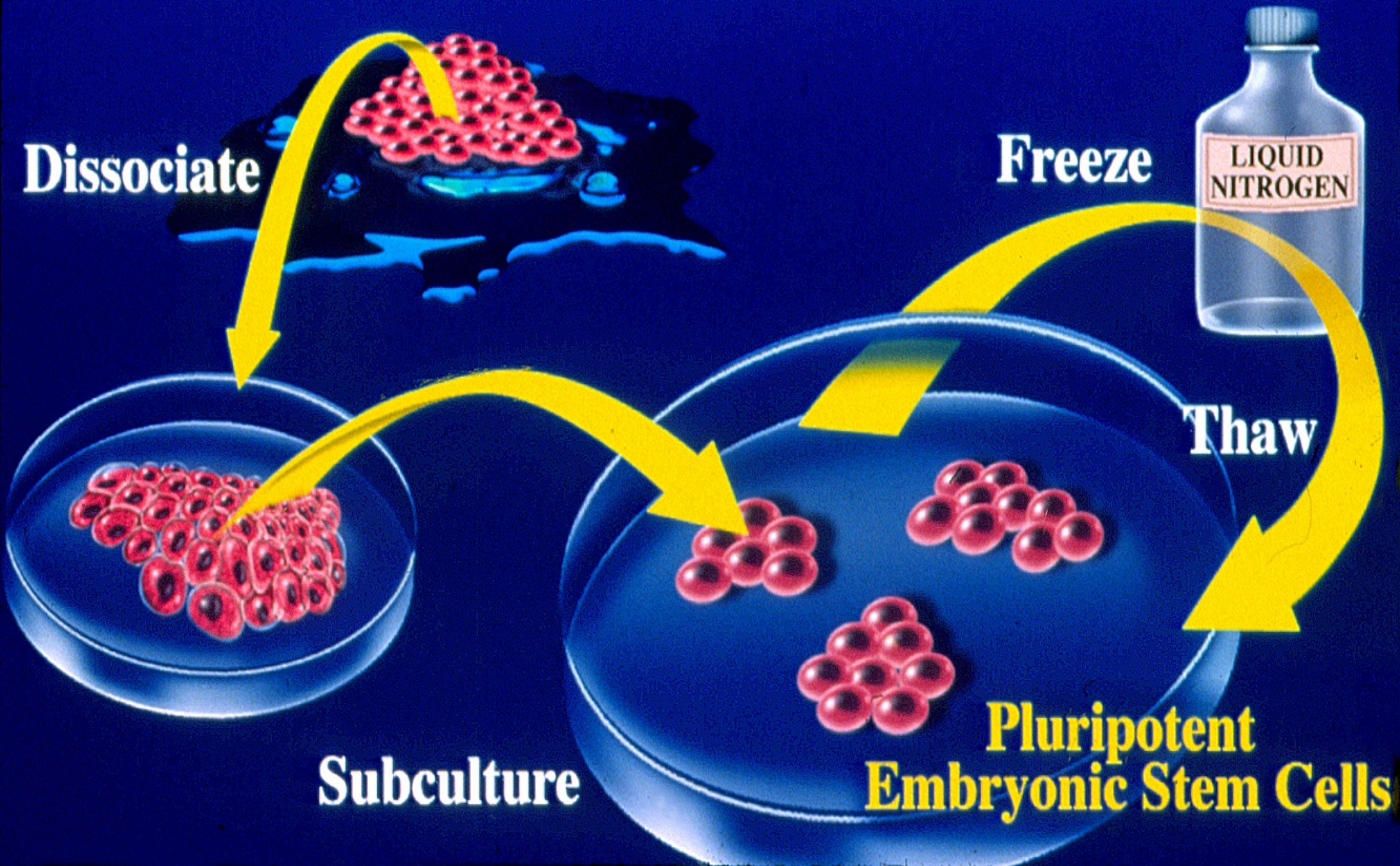


James A. Thomson, *Science* 282, 1145 (1998)

Courtesy Paul Berg

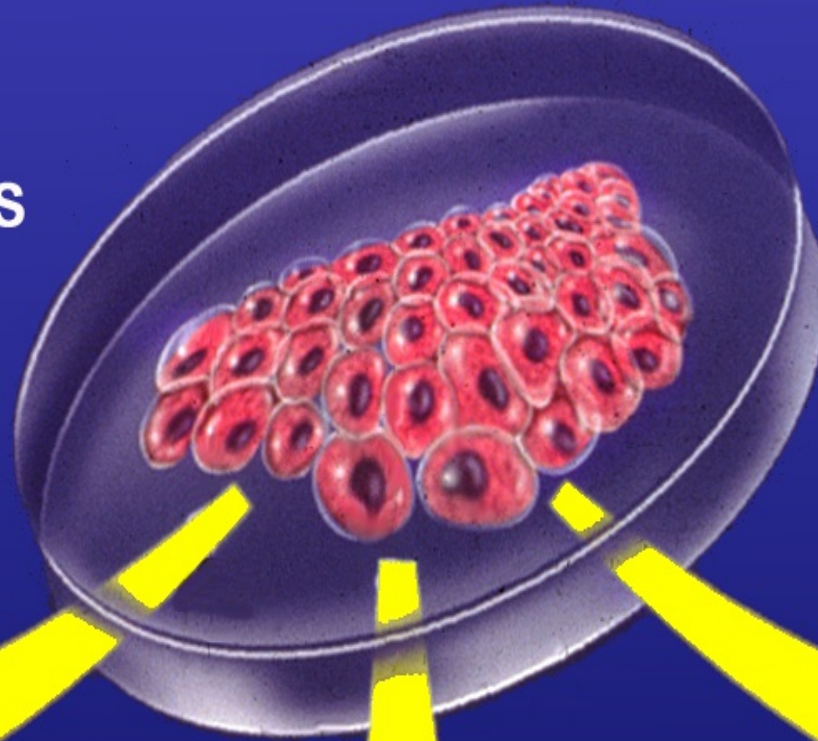


# Inner Cell Mass Cells Continue to Proliferate Indefinitely in Culture

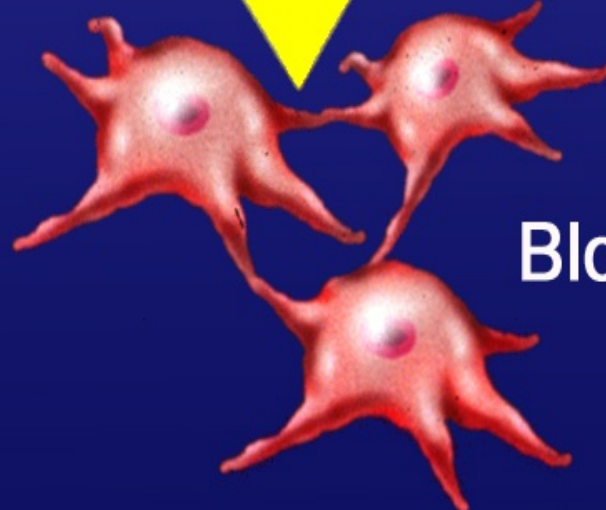


# Pluripotent Stem Cells Differentiate into many Cell Types

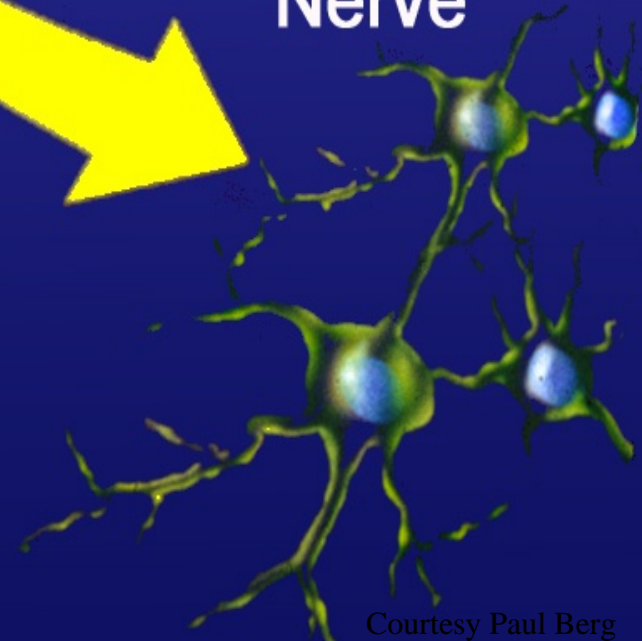
Add different growth factors



Muscle



Blood

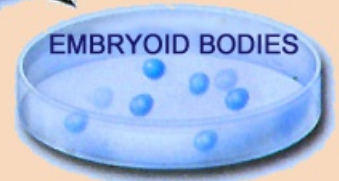
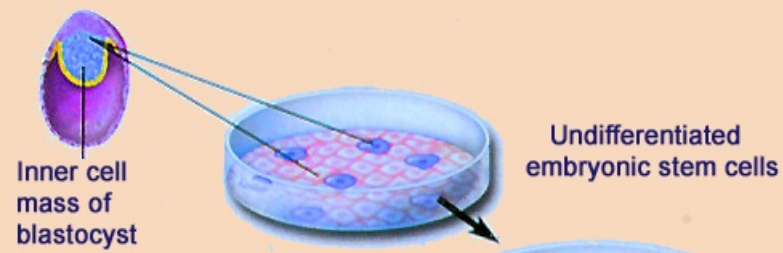


Nerve

# Basic Problems of Stem Cell Therapy

---

- HOW TO DIRECT DIFFERENTIATION OF CELLS DOWN SPECIFIC PATHWAYS?  
e.g. all into muscle or all into nerve; different “cocktails” of growth factors
- HOW TO OVERCOME IMMUNE REJECTION?  
e.g. alter histocompatibility genes; therapeutic cloning for “customized” lines
- HOW TO MAKE AN ORGAN?  
e.g. combine different cell types in three dimensional arrangements.



ITFSn medium (insulin/transferrin/  
fibronectin/selenium)

Adherent substrate

SELECTION OF NESTIN-POSITIVE CELLS

N2 medium/bFGF/laminin

N2 medium/bFGF/  
B27 media supplement

Expansion  
Phase

NESTIN-POSITIVE NEURONAL  
PRECURSOR CELLS

NESTIN-POSITIVE PANCREATIC  
PROGENITOR CELLS

Remove bFGF

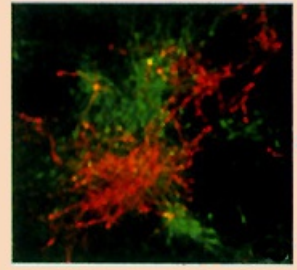
Differentiation  
Phase

Remove bFGF  
Add nicotinamide

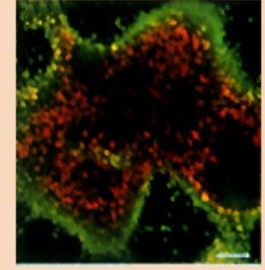
DOPAMINE- AND SERATONIN-  
SECRETING NEURONS

INSULIN-SECRETING PANCREATIC  
ISLET-LIKE CLUSTER

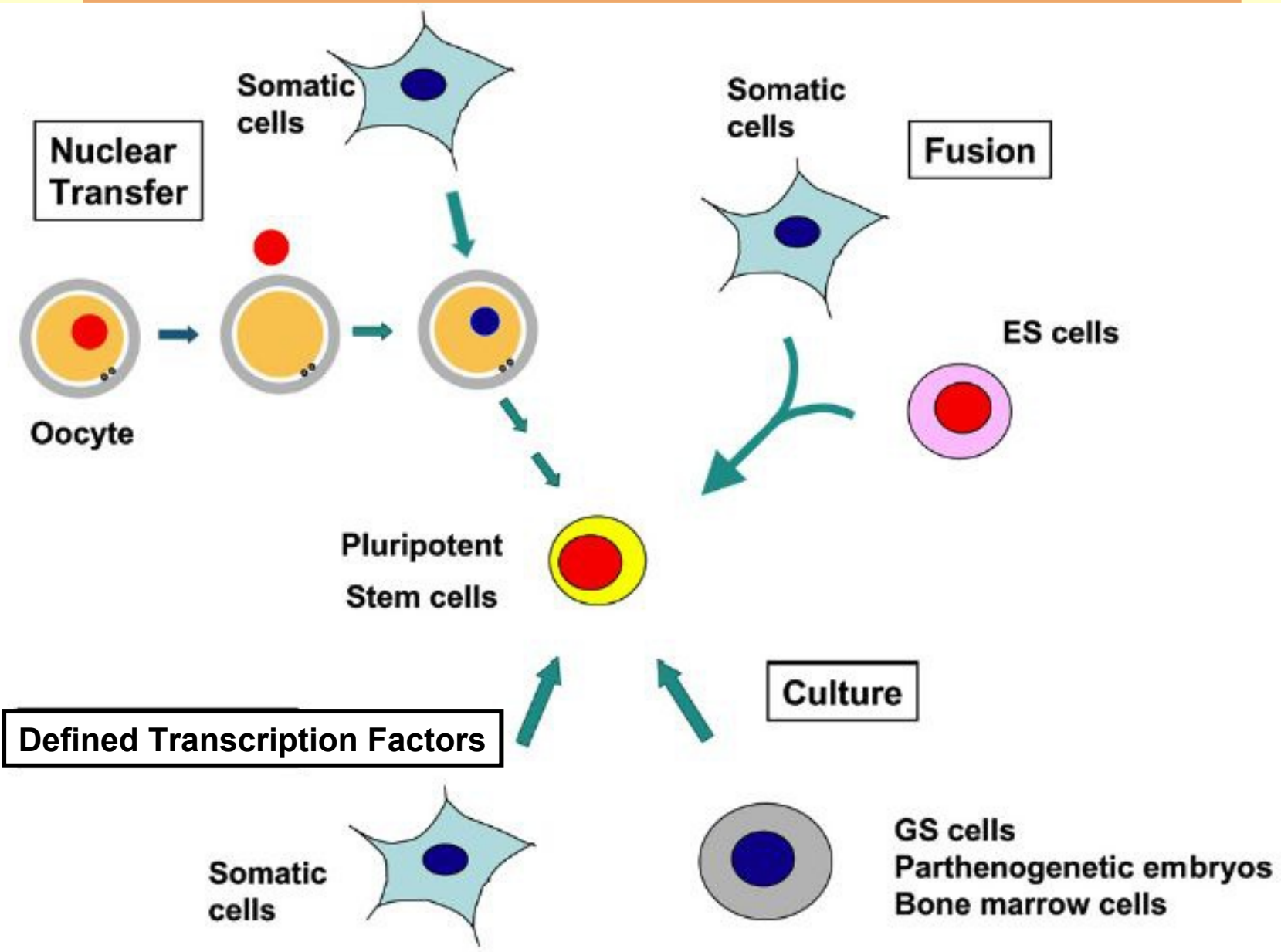
TYROSINE HYDROXYLASE/SEROTONIN



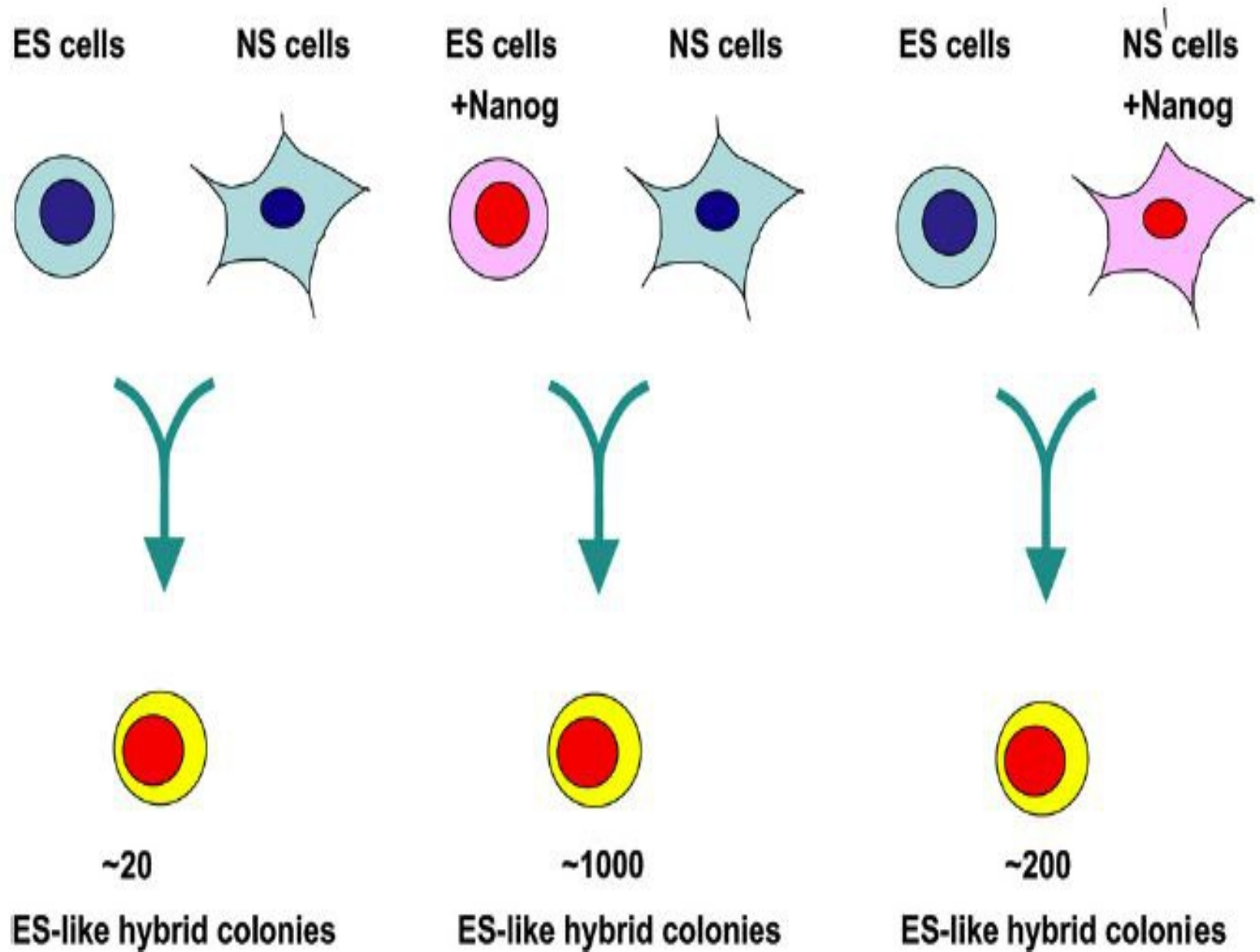
INSULIN/GLUCAGON



# Methods to Generate Pluripotent Stem Cells



# Nanog-Mediated Enhancement of Reprogramming by Fusion

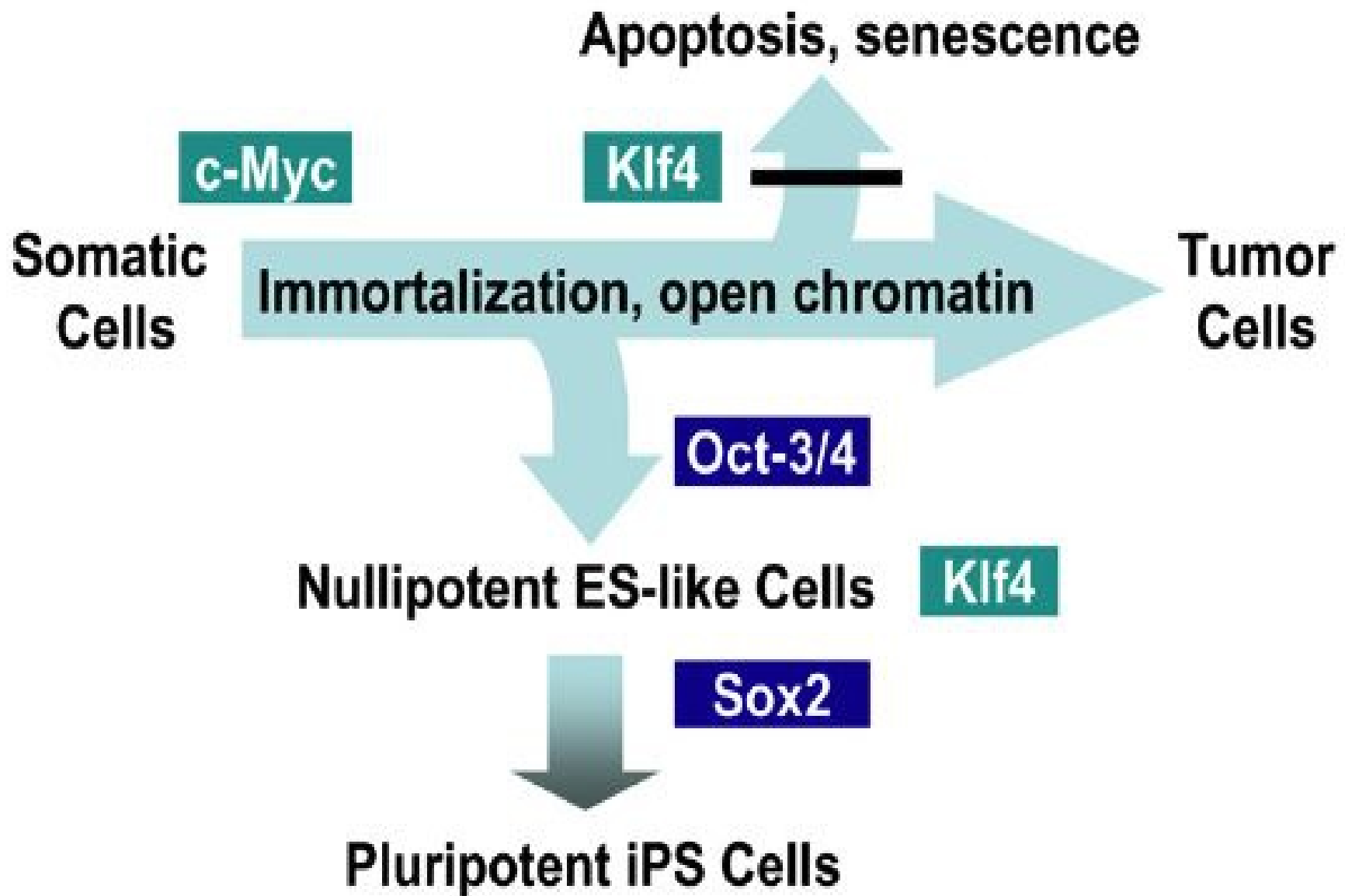


# Five Transcription Factors Needed to Maintain Pluripotency

**Table 1. Comparison of the Five Factors in the Phenotype of Loss-of-Function and Gain-of-Function Experiments**

	Knockout ES Cells	Knockout Embryos	Overexpression in ES Cells
<i>Oct-3/4</i>	Cannot be established	No epiblast	Induces differentiation
	Niwa et al., 2000	Nichols et al., 1998	Niwa et al., 2000
<i>Sox2</i>	Cannot be established	No epiblast	Does not induce differentiation
	Masui et al., 2007	Avilion et al., 2003	Does not induce LIF independency
			M. Nakagawa and S.Y., unpublished data
<i>c-Myc</i>	Can be established	Normal epiblast	Does not induce differentiation
	Normal self-renewal		Induces LIF independency
	Davis et al., 1993	Davis et al., 1993	Cartwright et al., 2005
<i>KLF4</i>	Not reported	Normal epiblast	Does not induce differentiation
		Katz et al., 2002	Induces LIF independency
			Y. Tokuzawa, M. Nakagawa, and S.Y., unpublished data
<i>Nanog</i>	Can be established	No epiblast	Does not induce differentiation
	Spontaneous differentiation		Induces LIF independency
	Mitsui et al., 2003	Mitsui et al., 2003	Chambers et al., 2003; Mitsui et al., 2003

# Induction of Pluripotent Stem Cells (iPS) from Somatic Stem Cells





# Adipose Tissue Provides iPSC Efficiently

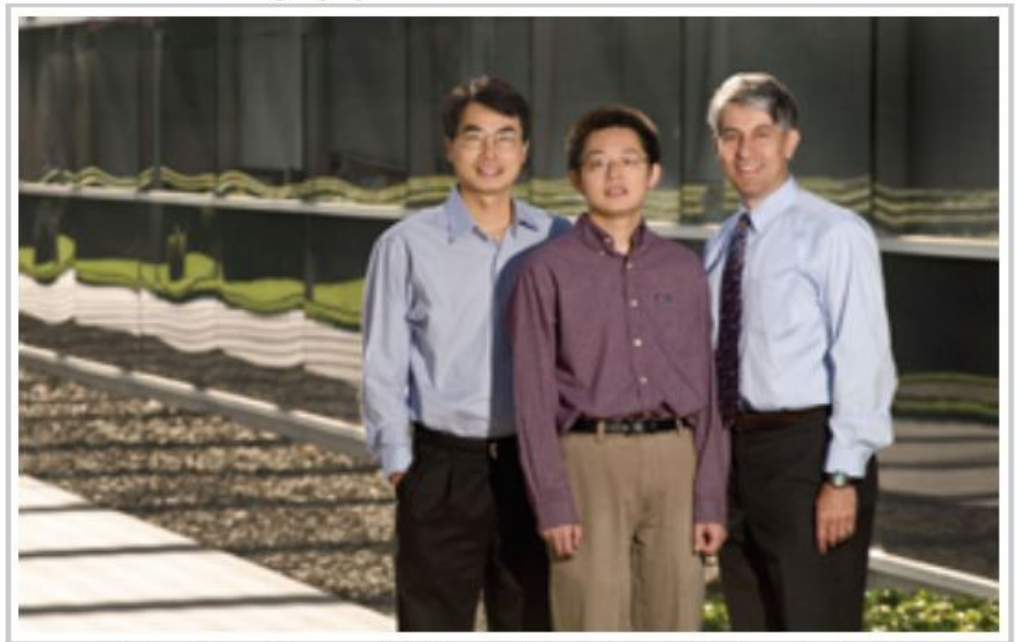
## 'Liposuction leftovers' easily converted to iPS cells, study shows

BY KRISTA CONGER

Globs of human fat removed during liposuction conceal versatile cells that are more quickly and easily coaxed to become induced pluripotent stem cells, or iPS cells, than are the skin cells most often used by researchers, according to a new study from Stanford's School of Medicine.

"We've identified a great natural resource," said Stanford surgery professor and co-author of the research, Michael Longaker, MD, who has called the readily available liposuction leftovers "liquid gold." Reprogramming adult cells to function like embryonic stem cells is one way researchers hope to create patient-specific cell lines to regenerate tissue or to study specific diseases in the laboratory.

Steve Fisch Photography



Joseph Wu, Ning Sun and Michael Longaker collaborated on research that showed stem cells found in fat tissue could easily be converted into iPS cells.

# Using CRE – Recombinase to Remove Viral Transforming DNA from iPSCs

---

## Parkinson's Disease Patient-Derived Induced Pluripotent Stem Cells Free of Viral Reprogramming Factors

Frank Soldner,<sup>1,4</sup> Dirk Hockemeyer,<sup>1,4</sup> Caroline Beard,<sup>1</sup> Qing Gao,<sup>1</sup> George W. Bell,<sup>1</sup> Elizabeth G. Cook,<sup>1</sup> Gunnar Hargus,<sup>3</sup> Alexandra Blak,<sup>3</sup> Oliver Cooper,<sup>3</sup> Maisam Mitalipova,<sup>1</sup> Ole Isacson,<sup>3</sup> and Rudolf Jaenisch<sup>1,2,\*</sup>

<sup>1</sup>The Whitehead Institute, 9 Cambridge Center, Cambridge, MA 02142, USA

<sup>2</sup>Department of Biology, Massachusetts Institute of Technology, 31 Ames Street, Cambridge, MA 02139, USA

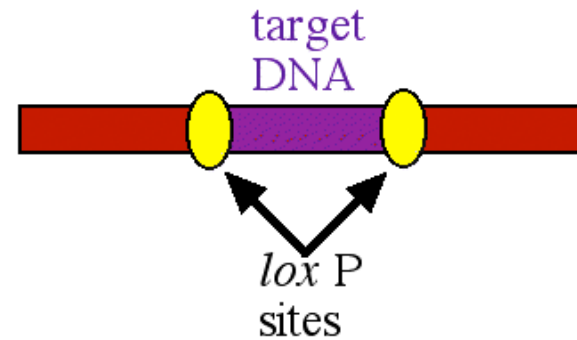
<sup>3</sup>Udall Parkinson Disease Research Center of Excellence, Center for Neurodegeneration Research, McLean Hospital/Harvard Medical School, Belmont, MA 02478, USA

<sup>4</sup>These authors contributed equally to this work

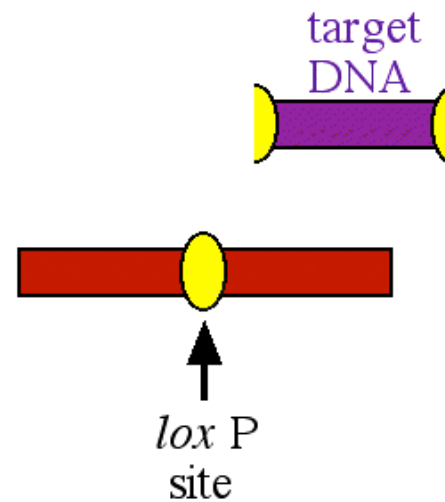
\*Correspondence: [jaenisch@wi.mit.edu](mailto:jaenisch@wi.mit.edu)

DOI 10.1016/j.cell.2009.02.013

# Cre-Lox Recombination to Remove Viral DNA



**Figure 1.** A pair of *lox P* sites (yellow ovals) flanking the target DNA (purple) to be deleted.



**Figure 2.** After the cre enzyme has excised the target DNA, one *lox P* site is left behind and the two flanking fragments of DNA are spliced together. The target DNA is excised and degraded.

# Inducing iPSCs using Transcription Factor Proteins

Cell  
PRESS

Cell Stem Cell  
Brief Report

## Generation of Human Induced Pluripotent Stem Cells by Direct Delivery of Reprogramming Proteins

Dohoon Kim,<sup>1,5</sup> Chun-Hyung Kim,<sup>1,5</sup> Jung-Il Moon,<sup>1</sup> Young-Gie Chung,<sup>3</sup> Mi-Yoon Chang,<sup>1</sup> Baek-Soo Han,<sup>1</sup> Sanghyeok Ko,<sup>1</sup> Eungi Yang,<sup>1</sup> Kwang Yul Cha,<sup>4</sup> Robert Lanza,<sup>3,\*</sup> and Kwang-Soo Kim<sup>1,2,4,\*</sup>

<sup>1</sup>Molecular Neurobiology Laboratory, Department of Psychiatry and McLean Hospital, Harvard Medical School

<sup>2</sup>Harvard Stem Cell Institute

115 Mill Street, Belmont, MA 02478, USA

<sup>3</sup>Stem Cell and Regenerative Medicine International, 381 Plantation Street, Worcester, MA 01605, USA

<sup>4</sup>CHA Stem Cell Institute, CHA University, 606-16 Yoeksam 1-dong, Gangnam-gu, Korea

<sup>5</sup>These authors contributed equally to this work


\*Correspondence: [rlanza@advancedcell.com](mailto:rlanza@advancedcell.com) (R.L.), [kskim@mclean.harvard.edu](mailto:kskim@mclean.harvard.edu) (K.-S.K.)

DOI 10.1016/j.stem.2009.05.005

# Direct conversion of mouse fibroblasts to self-renewing, tripotent neural precursor cells

Ernesto Lujan<sup>a,b</sup>, Soham Chanda<sup>a,c</sup>, Henrik Ahlenius<sup>a,d</sup>, Thomas C. Südhof<sup>c,e,1</sup>, and Marius Wernig<sup>a,d,1</sup>

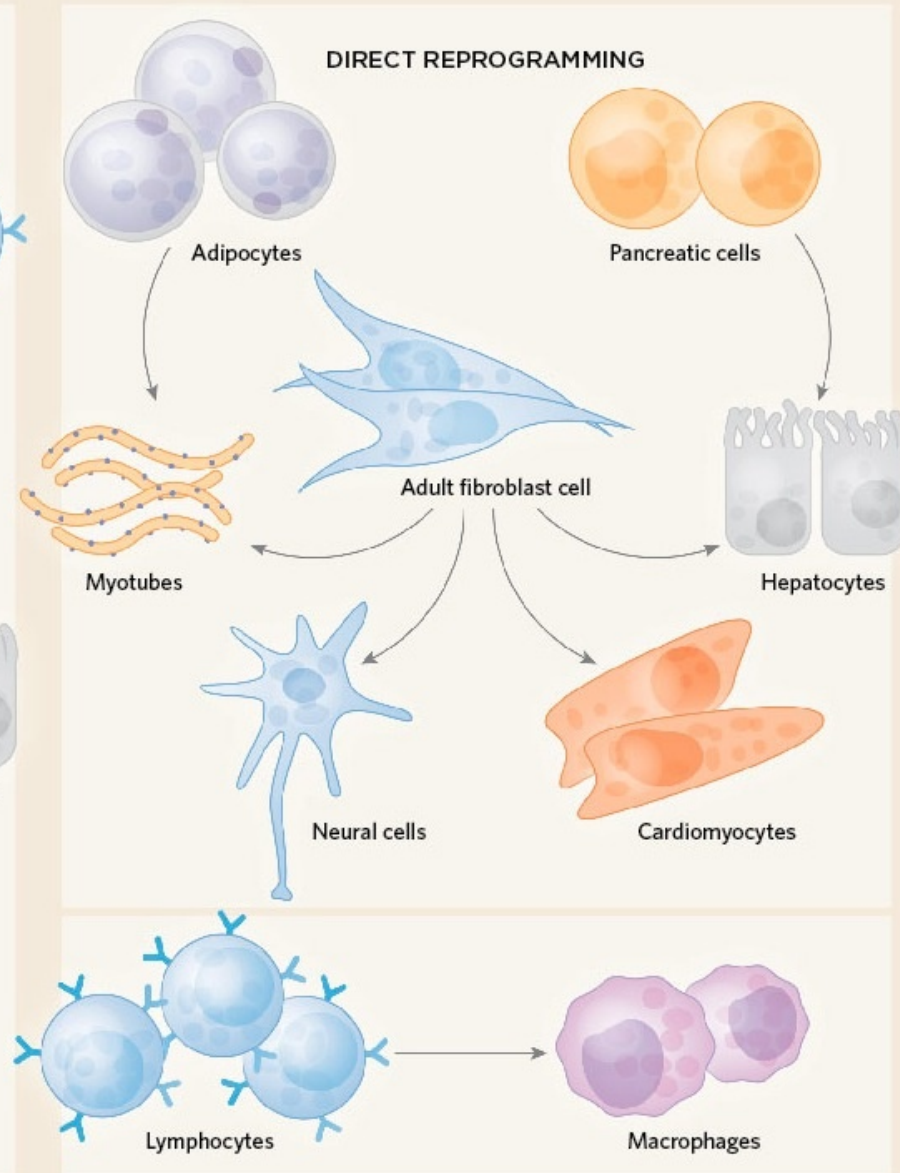
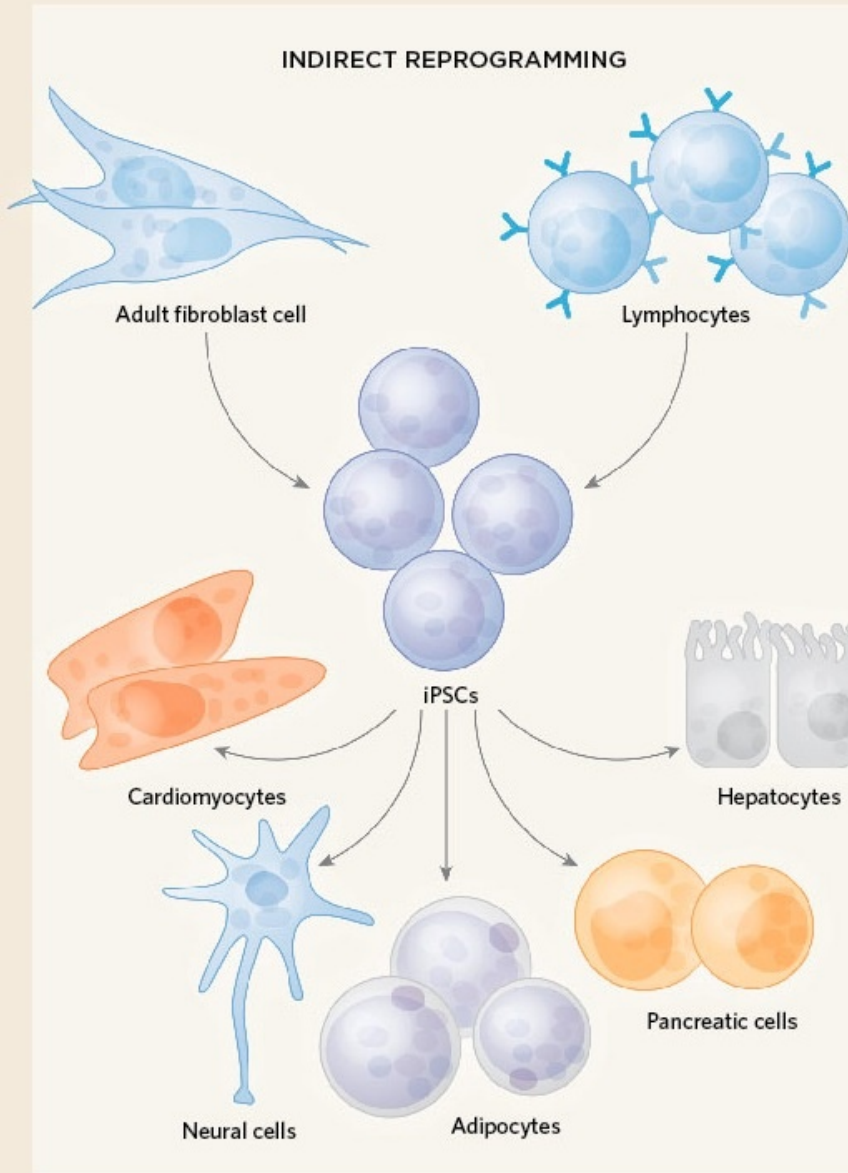
<sup>a</sup>Institute for Stem Cell Biology and Regenerative Medicine, Departments of <sup>d</sup>Pathology, <sup>b</sup>Genetics, and <sup>c</sup>Molecular and Cellular Physiology, and <sup>e</sup>Howard Hughes Medical Institute, Stanford University School of Medicine, Stanford, CA 94305



We recently showed that defined sets of transcription factors are sufficient to convert mouse and human fibroblasts directly into cells resembling functional neurons, referred to as “induced neuronal” (iN) cells. For some applications however, it would be desirable to convert fibroblasts into proliferative neural precursor cells (NPCs) instead of neurons. We hypothesized that NPC-like cells may be induced using the same principal approach used for generating iN cells. Toward this goal, we infected mouse embryonic fibroblasts derived from Sox2-EGFP mice with a set of 11 transcription factors highly expressed in NPCs. Twenty-four days after transgene induction, Sox2-EGFP<sup>+</sup> colonies emerged that expressed NPC-specific genes and differentiated into neuronal and astrocytic cells. Using stepwise elimination, we found that Sox2 and FoxG1 are capable of generating clonal self-renewing, bipotent induced NPCs that gave rise to astrocytes and functional neurons. When we added the Pou and Homeobox domain-containing transcription factor **Brn2 to Sox2 and FoxG1, we were able to induce tripotent NPCs that could be differentiated not only into neurons and astrocytes but also into oligodendrocytes.** The transcription factors FoxG1 and Brn2 alone also were capable of inducing NPC-like cells; however, these cells generated less mature neurons, although they did produce astrocytes and even oligodendrocytes capable of integration into dysmyelinated Shiverer brain. Our data demonstrate that direct lineage reprogramming using target cell-type-specific transcription factors can be used to induce NPC-like cells that potentially could be used

# Direct Cell Reprogramming *in vivo* & *in vitro*

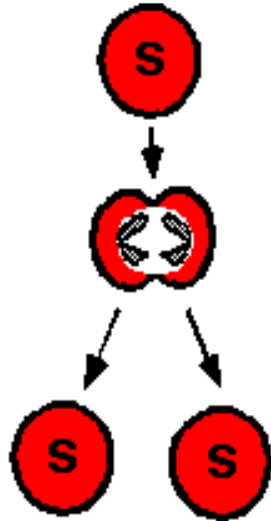
<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>



<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>

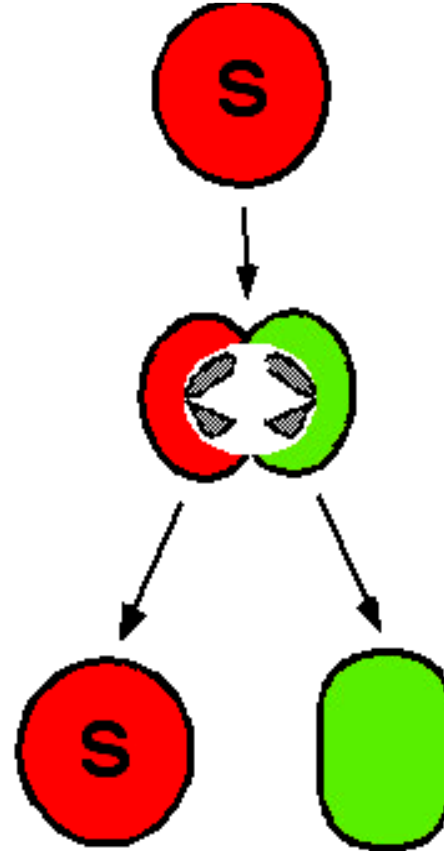
# Alternate Stem Cell Fates

Embryonic  
Stem Cells



stem cell  
proliferation

Adult  
Stem Cells



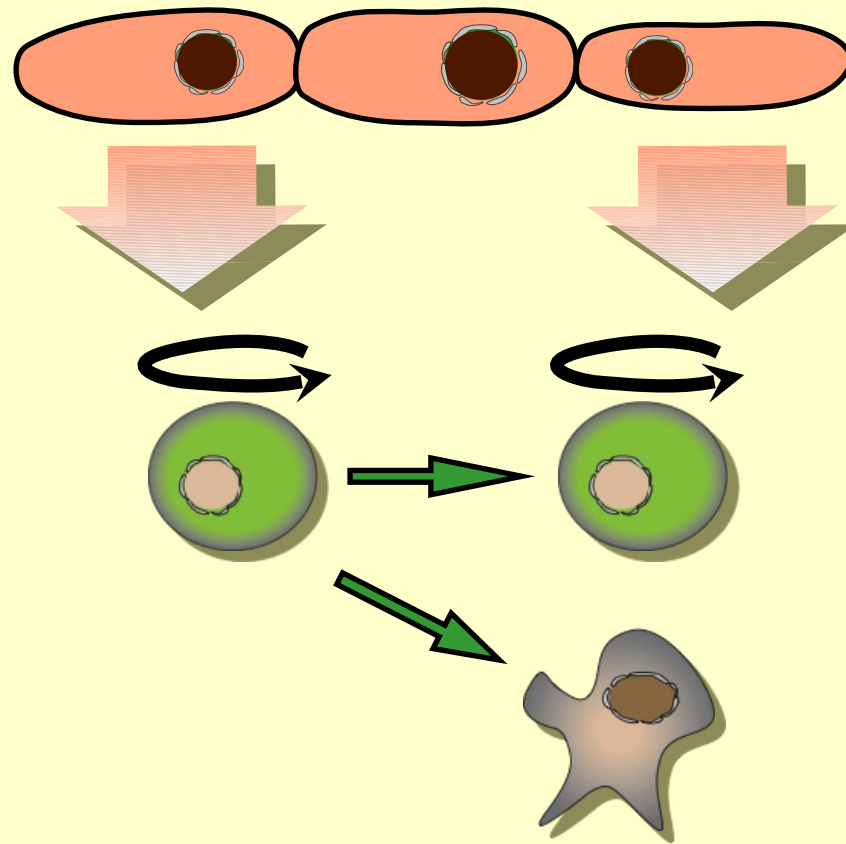
asymmetric  
division

Adult  
Stem Cells



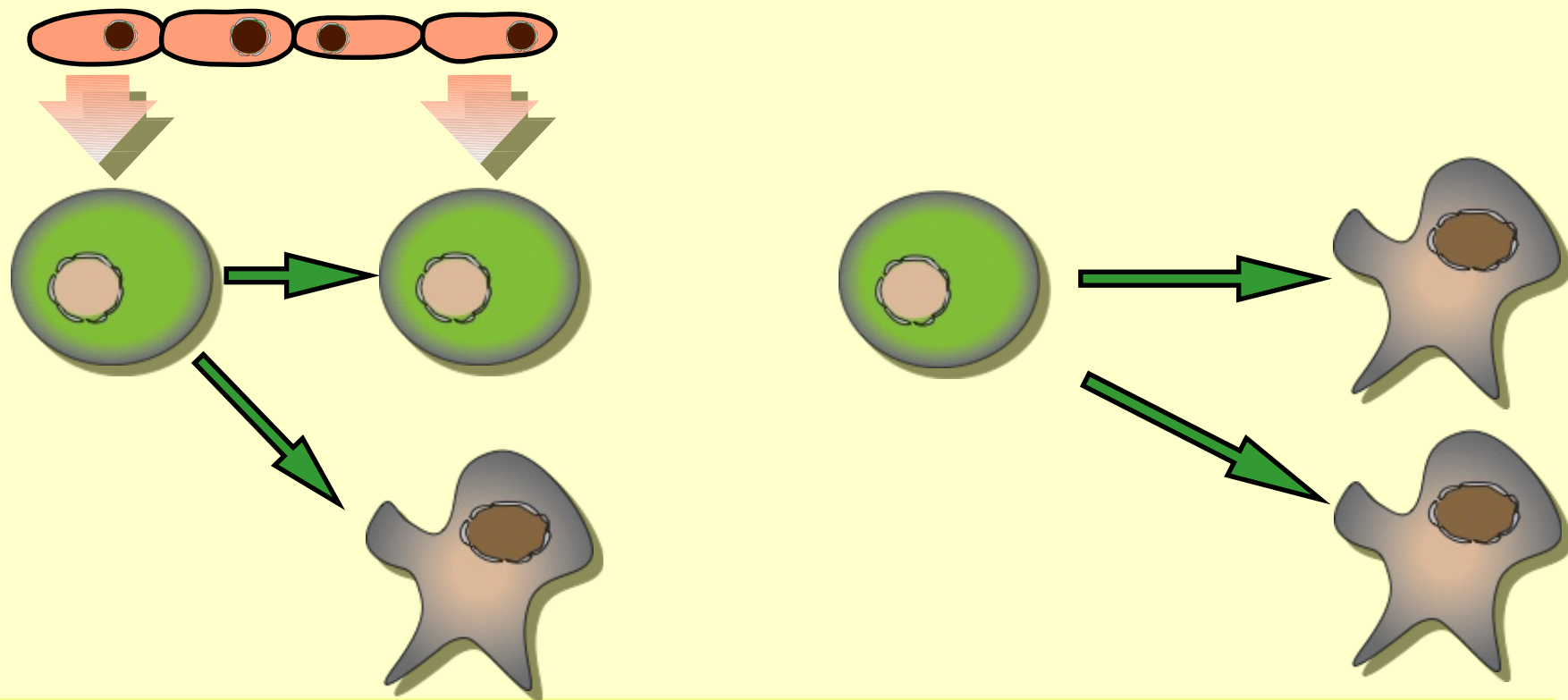
stem cell  
loss

# signals from niches maintain adult stem cells and tissues



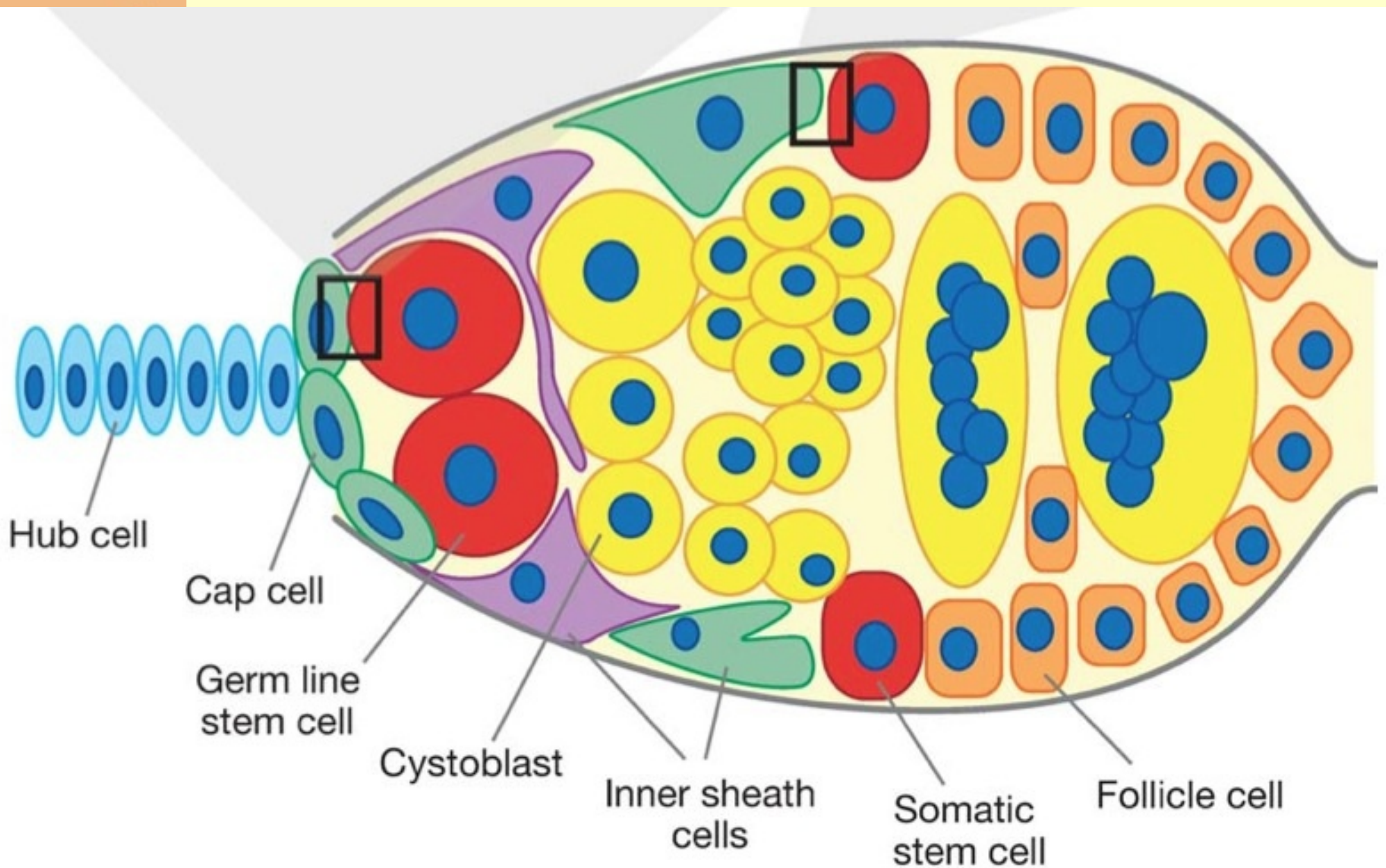


In the absence of niche signals, adult stem cells will differentiate, by default

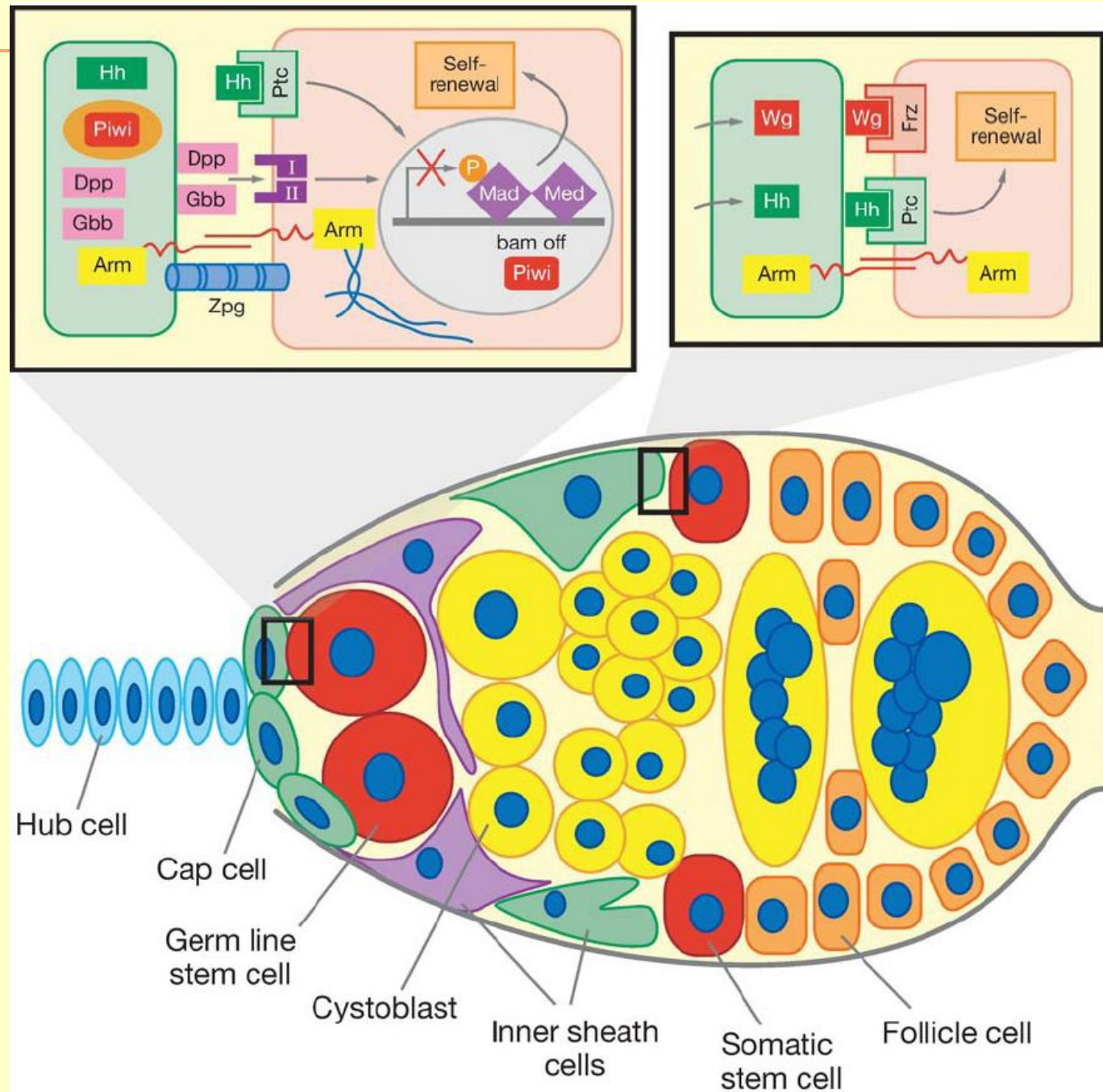


1. Self-renewal is proliferation coupled to blocking differentiation, controlled by signals.
2. Signals are local; niches have a limited capacity and cells compete for the signals
3. The signals control tissue homeostasis, also after damage

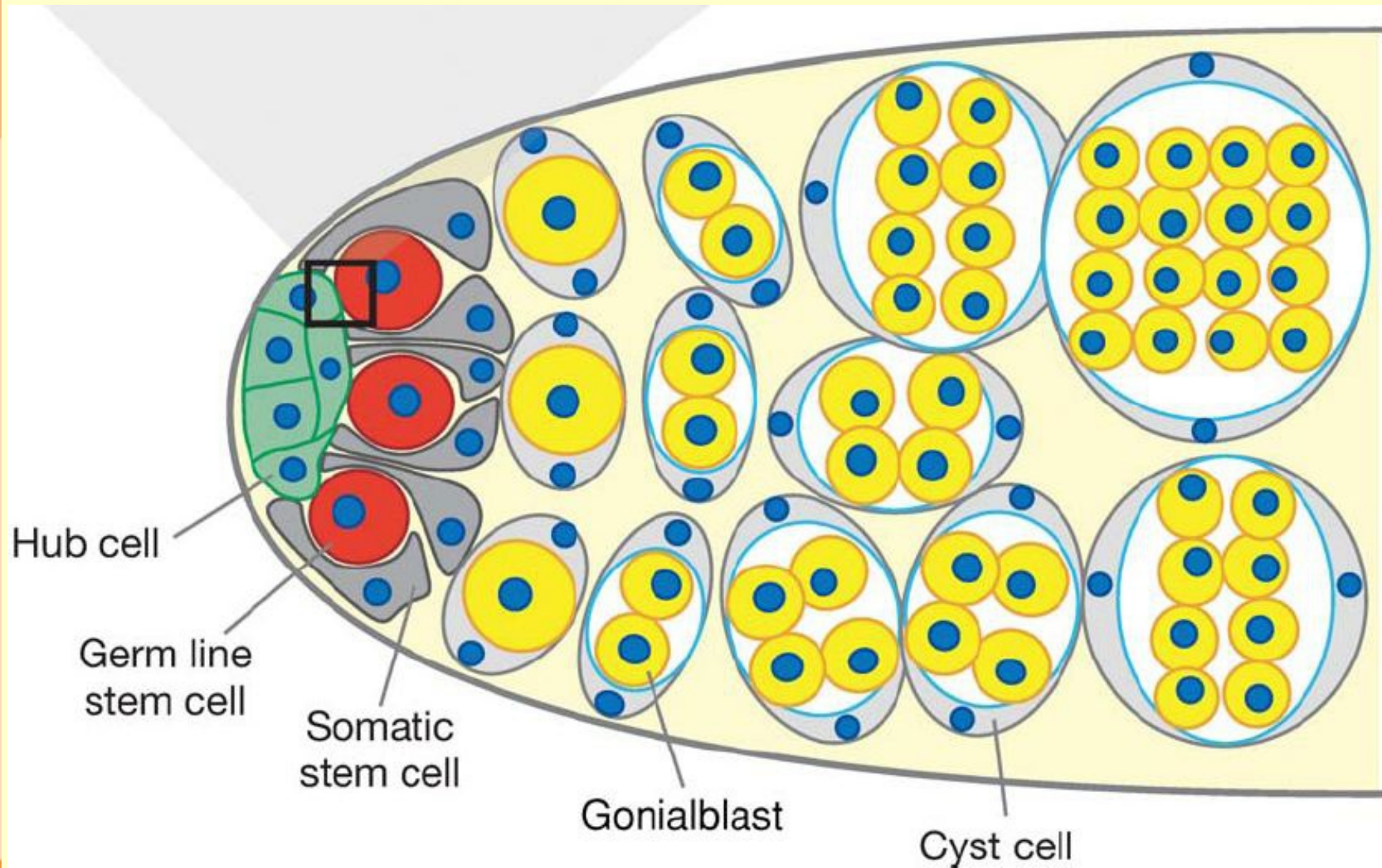
# Drosophila Oocyte Stem Cells



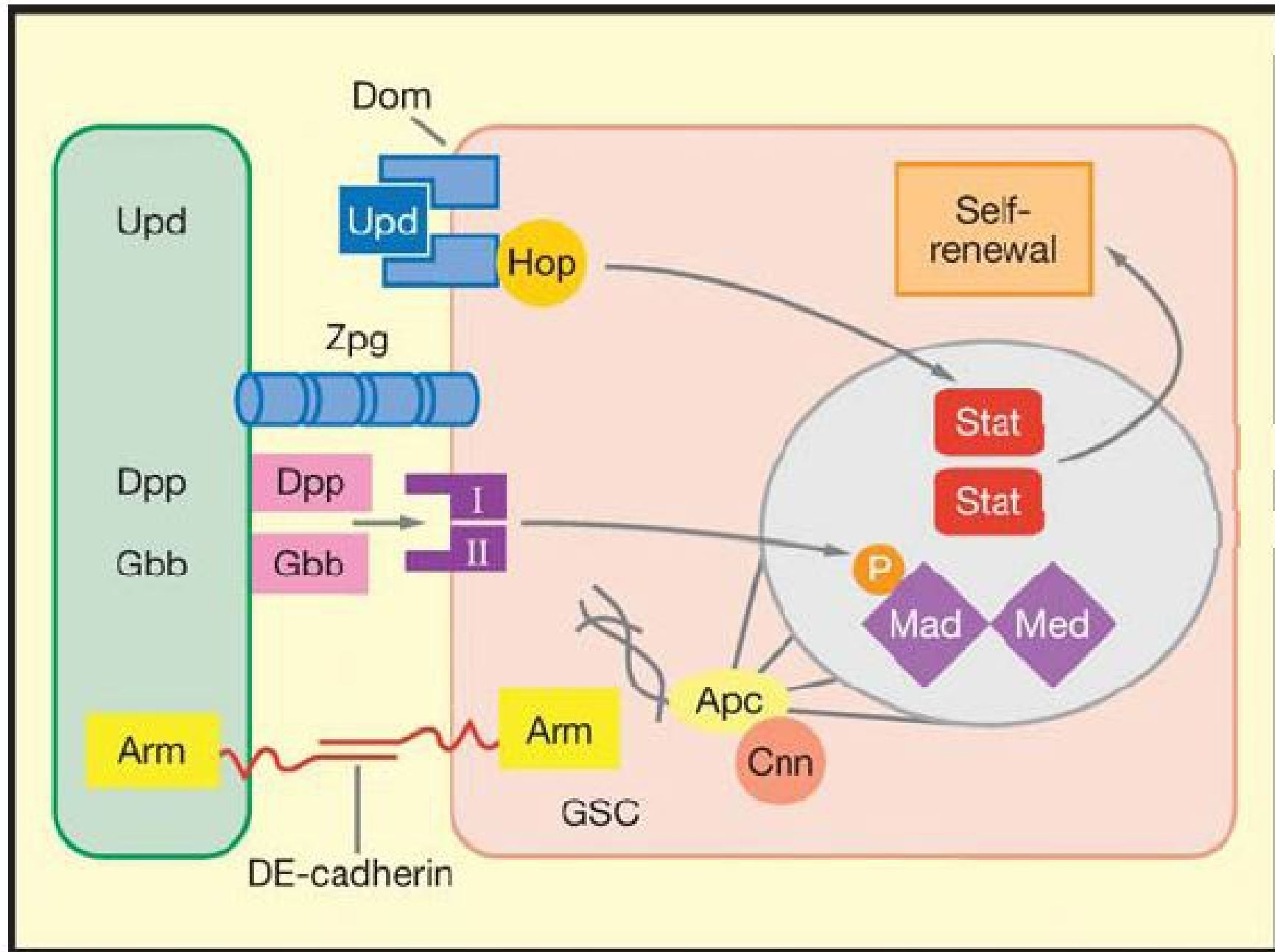
# Cell-Cell Interactions at Oocyte Niche



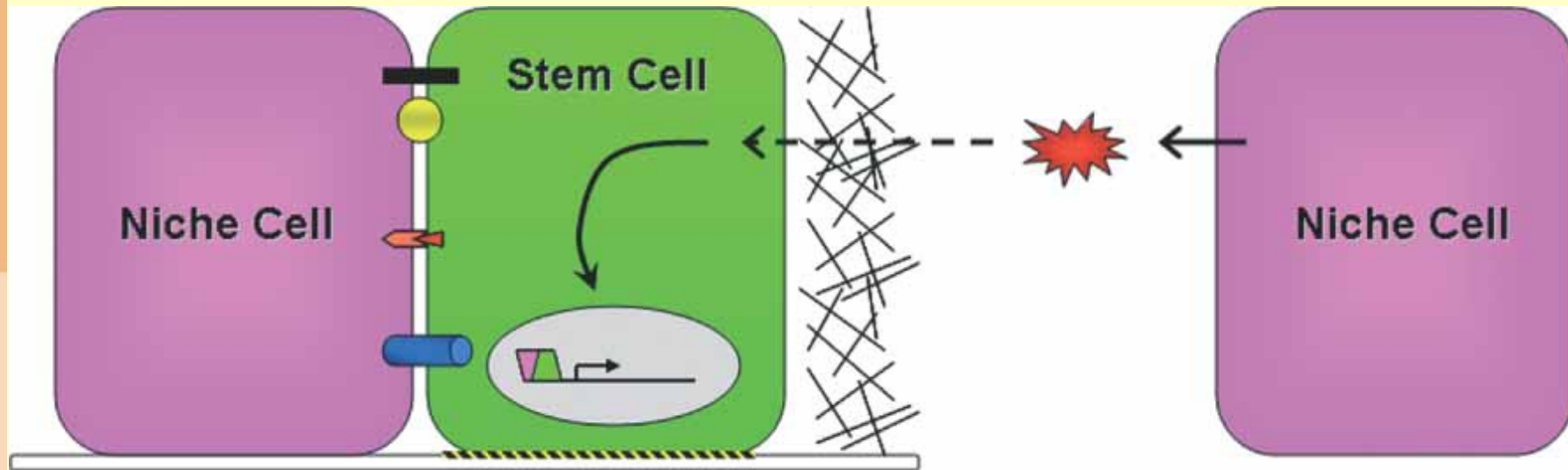
# Drosophila Spermatogonial Niche









# Cell-Cell Interactions at the Spermatogonial Niche






# Summary of Stem Cell Niche Signals



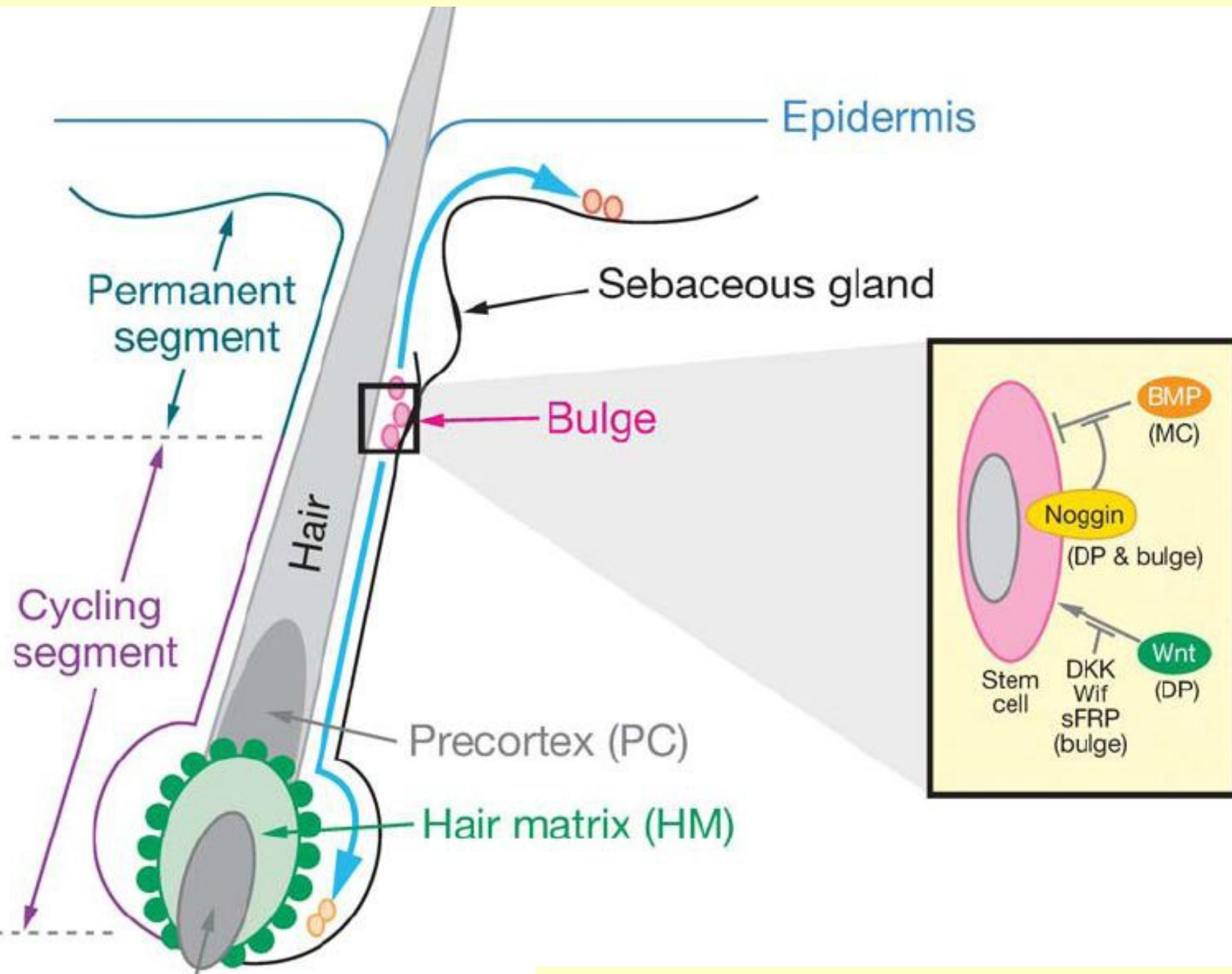
## Physical Contact

-  **Tight Junction**  
N, I
-  **Adherens Junction**  
D, N
-  **Notch Signaling**  
C, N, H, I
-  **Gap Junction**  
D
-  **Basement Membrane**  
N, E, I
-  **Extracellular Matrix**  
D, N,

## Diffusible Factors

-  **Pathway**  
Wnt: C, E, H, I  
BMP: D, N, E, I  
JAK/STAT: D  
Growth Factors: N  
Hedgehog: I  
PGE2: I  
O<sub>2</sub>: H
-  **Transcription Factor Activation**
-  **Signal Transduction**

# Hair Follicle Niche



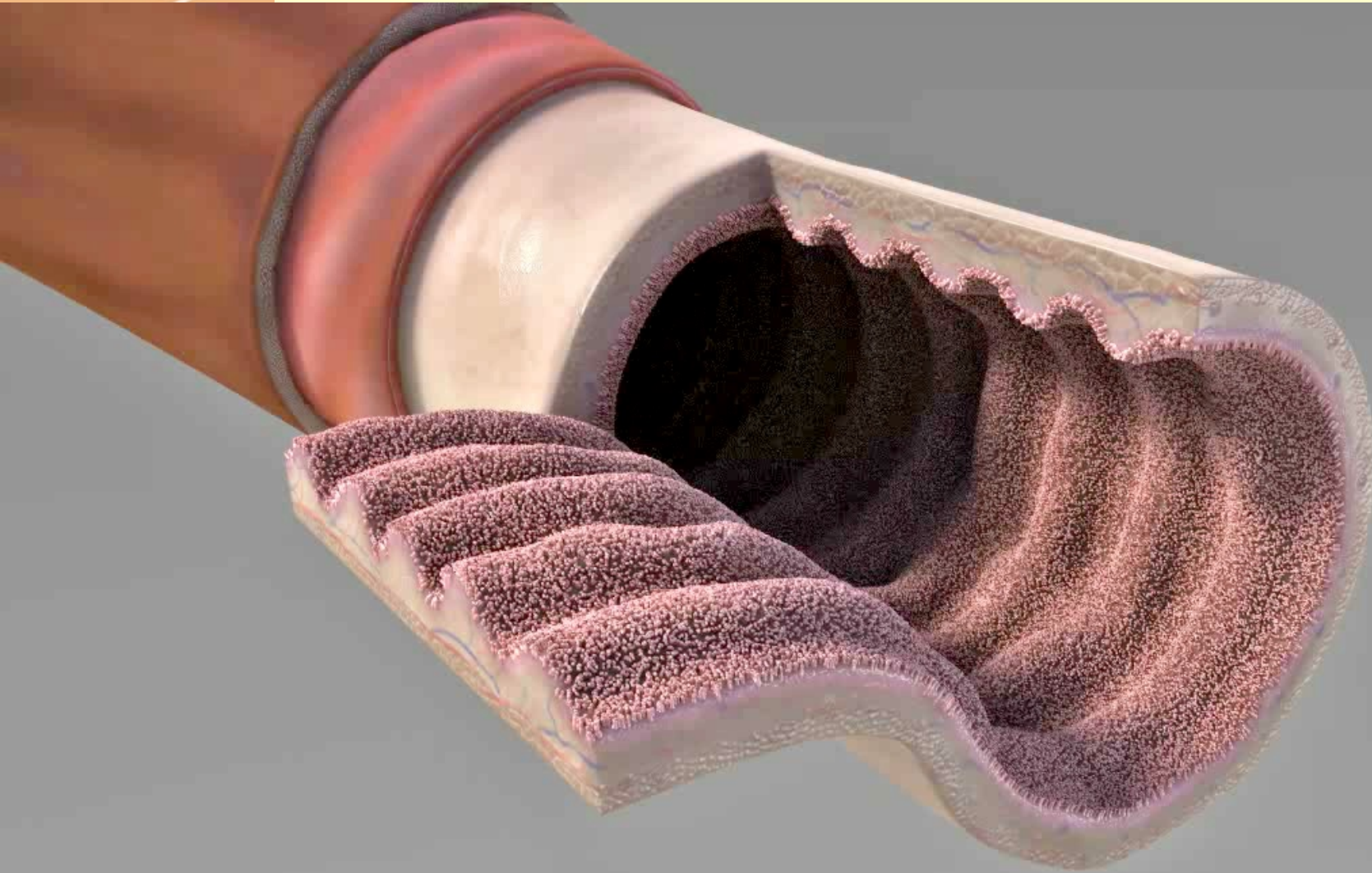
# Intestinal Stem Cells in Crypts

**Clevers Lab|Digizyme**





# Rainbow Villi

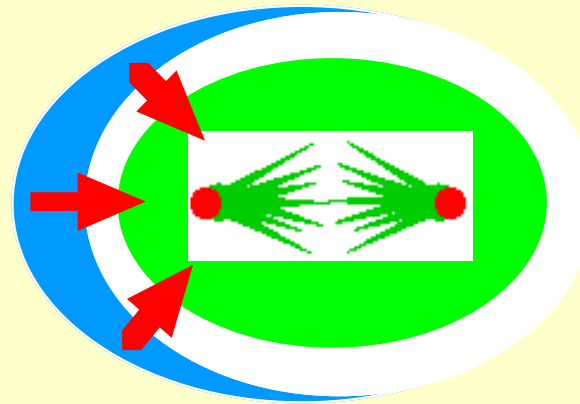


Clevers Lab | ANATOMY 3D

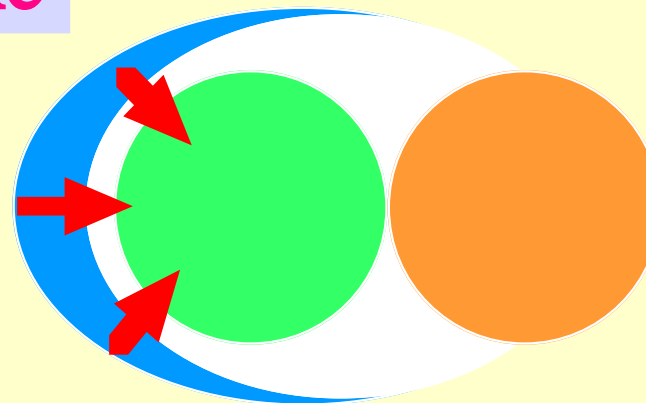
# Asymmetric stem cell divisions

---

Extrinsic  
factor(s)



Niche



# John Cairns: The Immortal Parental Strands

---

*Nature* Vol. 255 May 15 1975

197

---

## review article

---

### **Mutation selection and the natural history of cancer**

**John Cairns\***

---

*Survival of the rapidly renewing tissues of long-lived animals like man requires that they be protected against the natural selection of fitter variant cells (that is, the spontaneous appearance of cancer). This article discusses three possible protective mechanisms and shows how they could explain various features of the natural history of certain common cancers of man.*

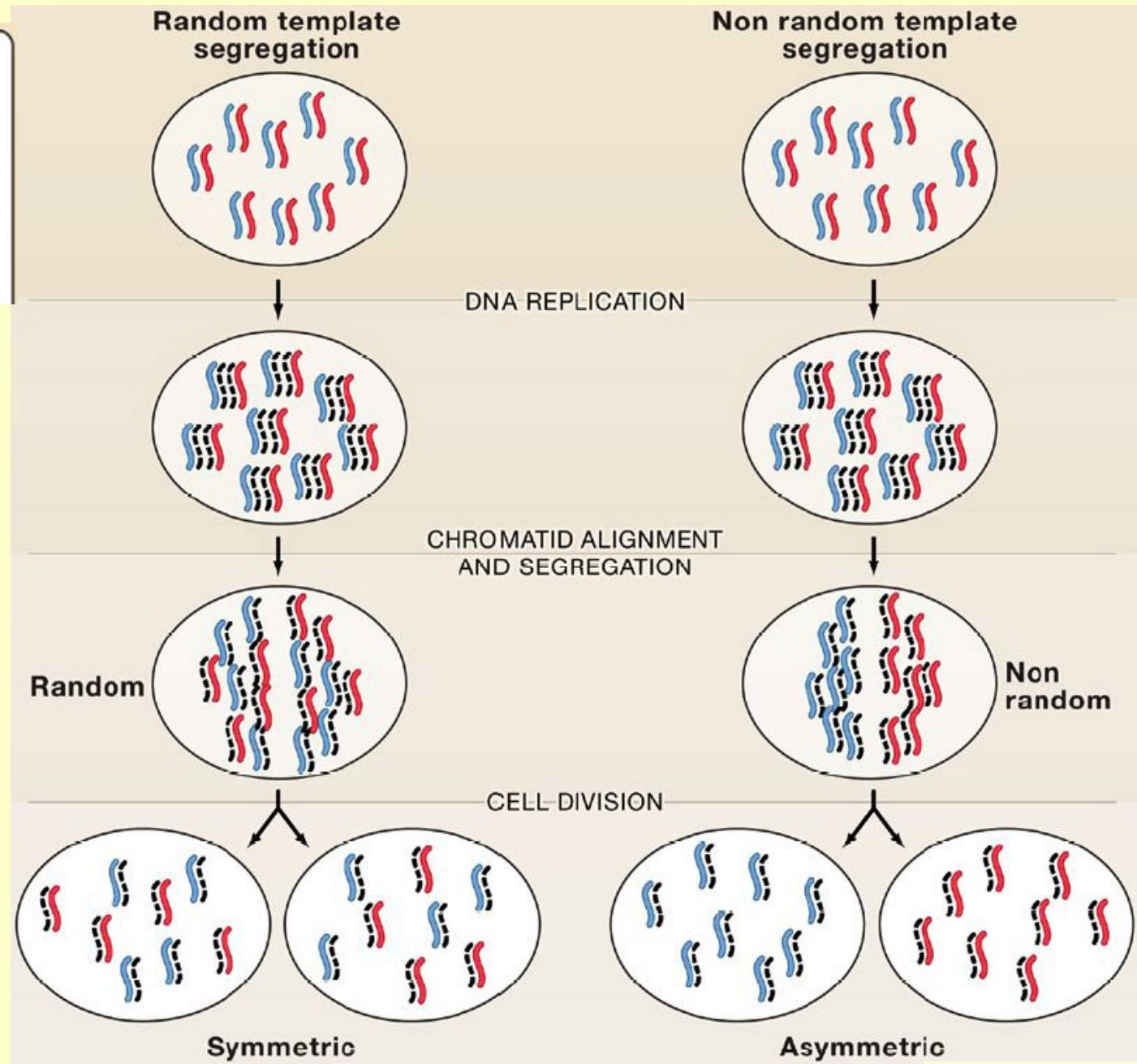
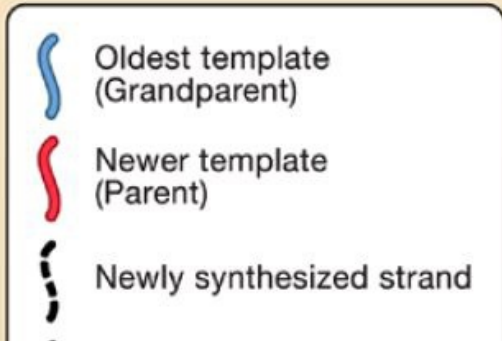
---

# Motivation for Asymmetric Strand Segregation

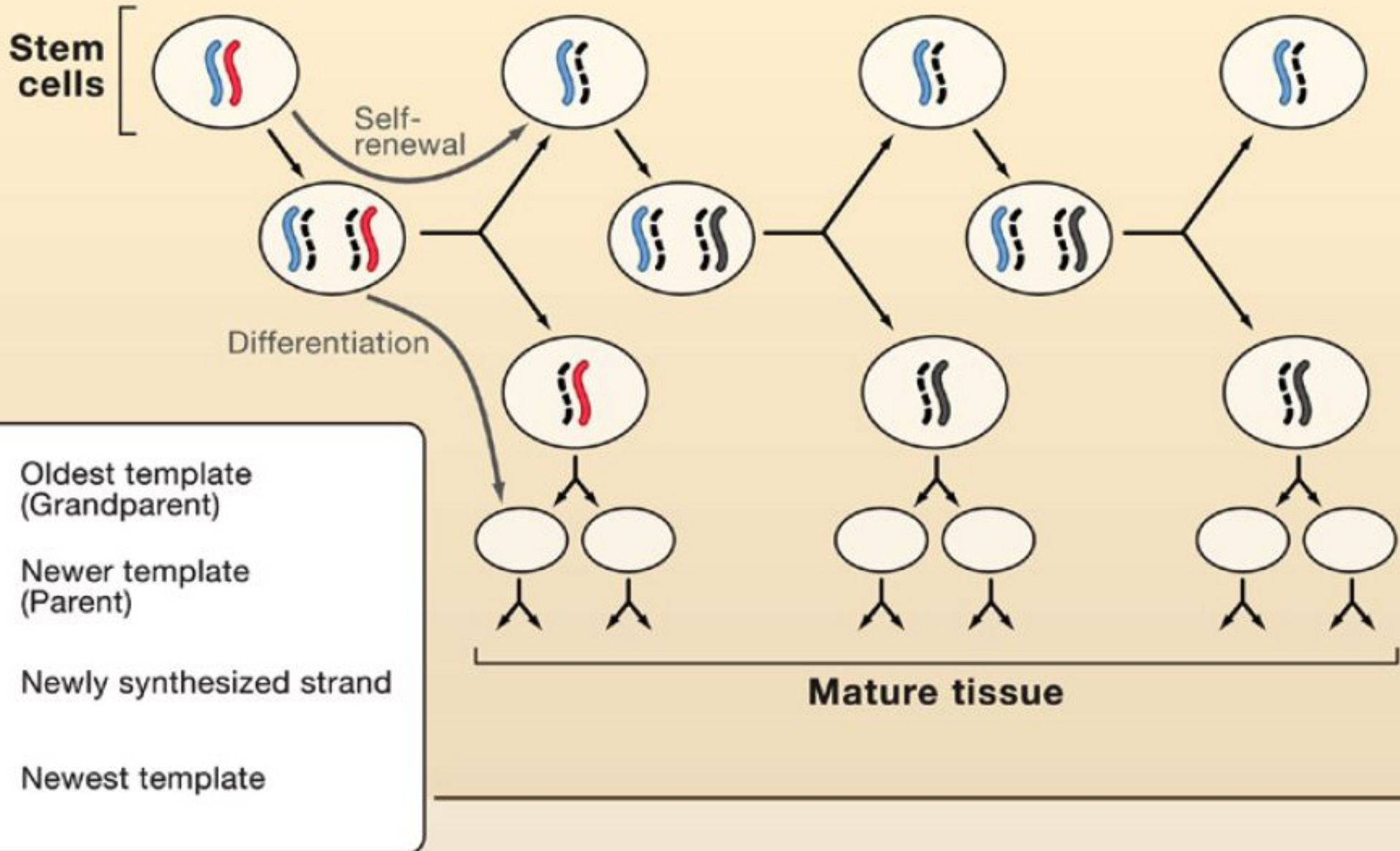
---

- Adult rat contains  $6 \times 10^{10}$  cells
- In its small intestine, a rat sheds over  $10^{13}$  epithelial cells during its lifetime.
- Requires  $10^3$  symmetric cell doublings from embryo to adult followed by  $10^{13}$  asymmetric cell doublings during its lifetime
- How do epithelial cells minimize mutations that lead to cancer?

# Asymmetric Segregation of Parental DNA Strands

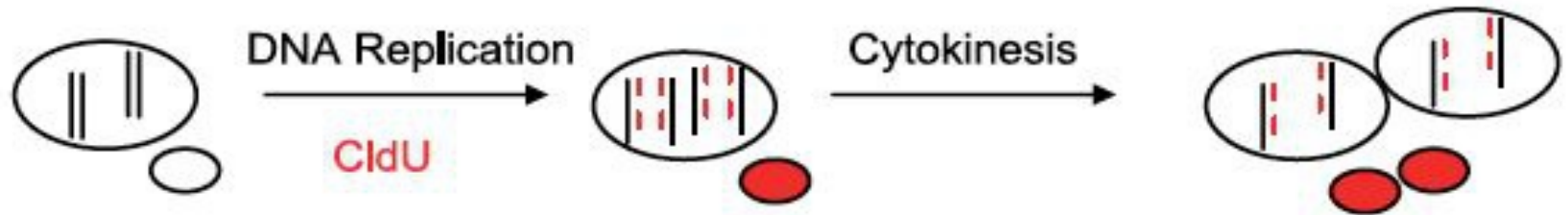


# Asymmetric Stem Cell Growth with Asymmetric Parental Strand Segregation

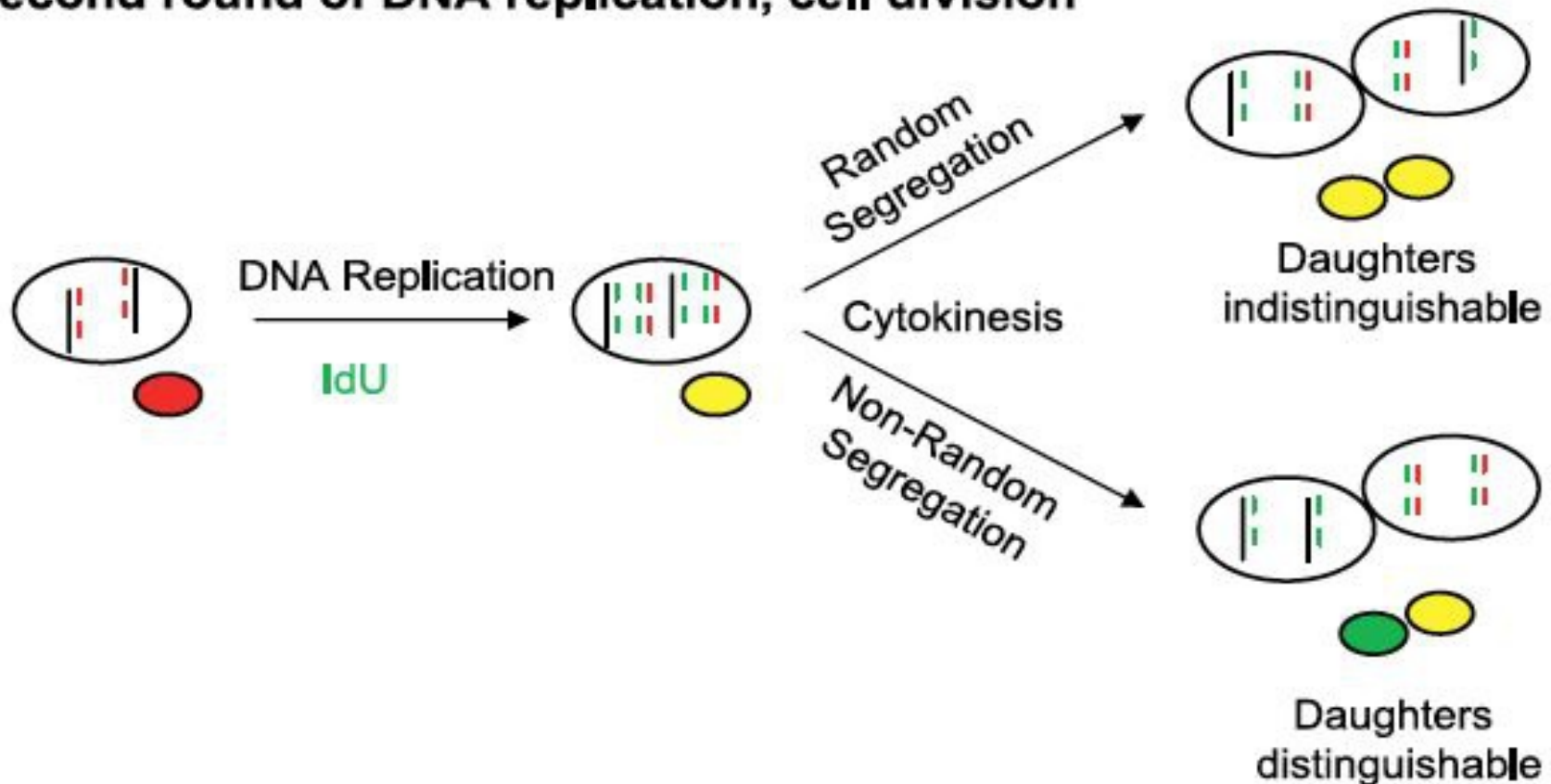


# Asymmetric DNA Labeling Patterns

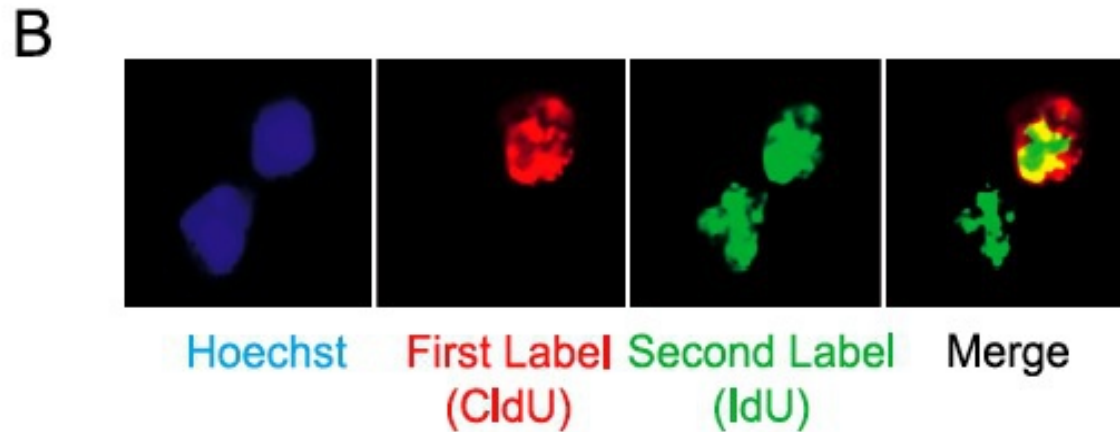
## First round of DNA replication, cell division



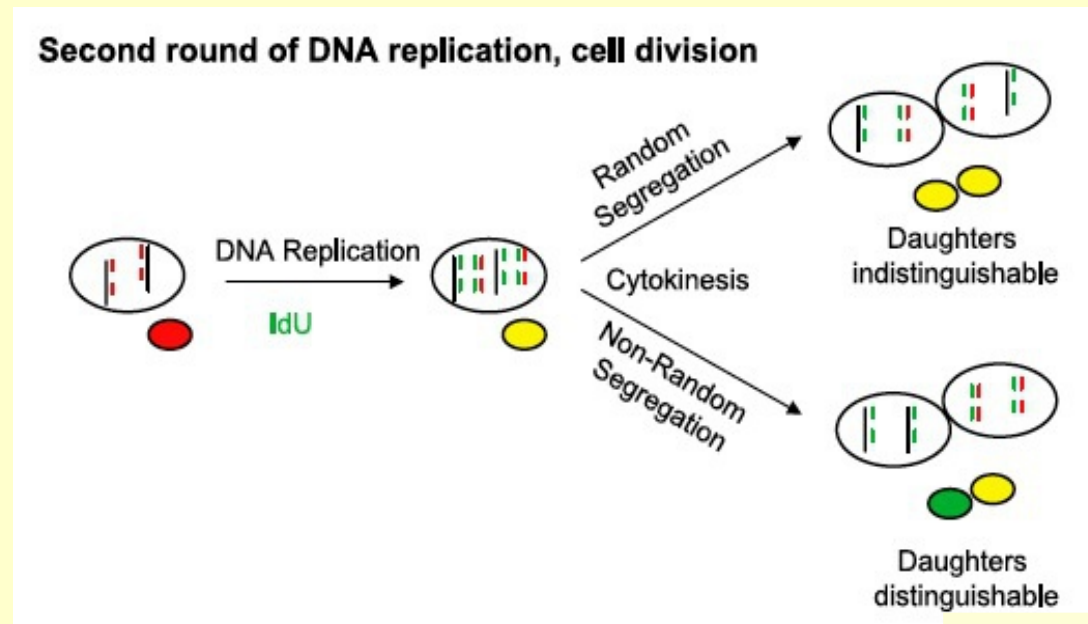
## Second round of DNA replication, cell division



# Duplicating Muscle Cell Pairs Display Asymmetric DNA Labeling Patterns

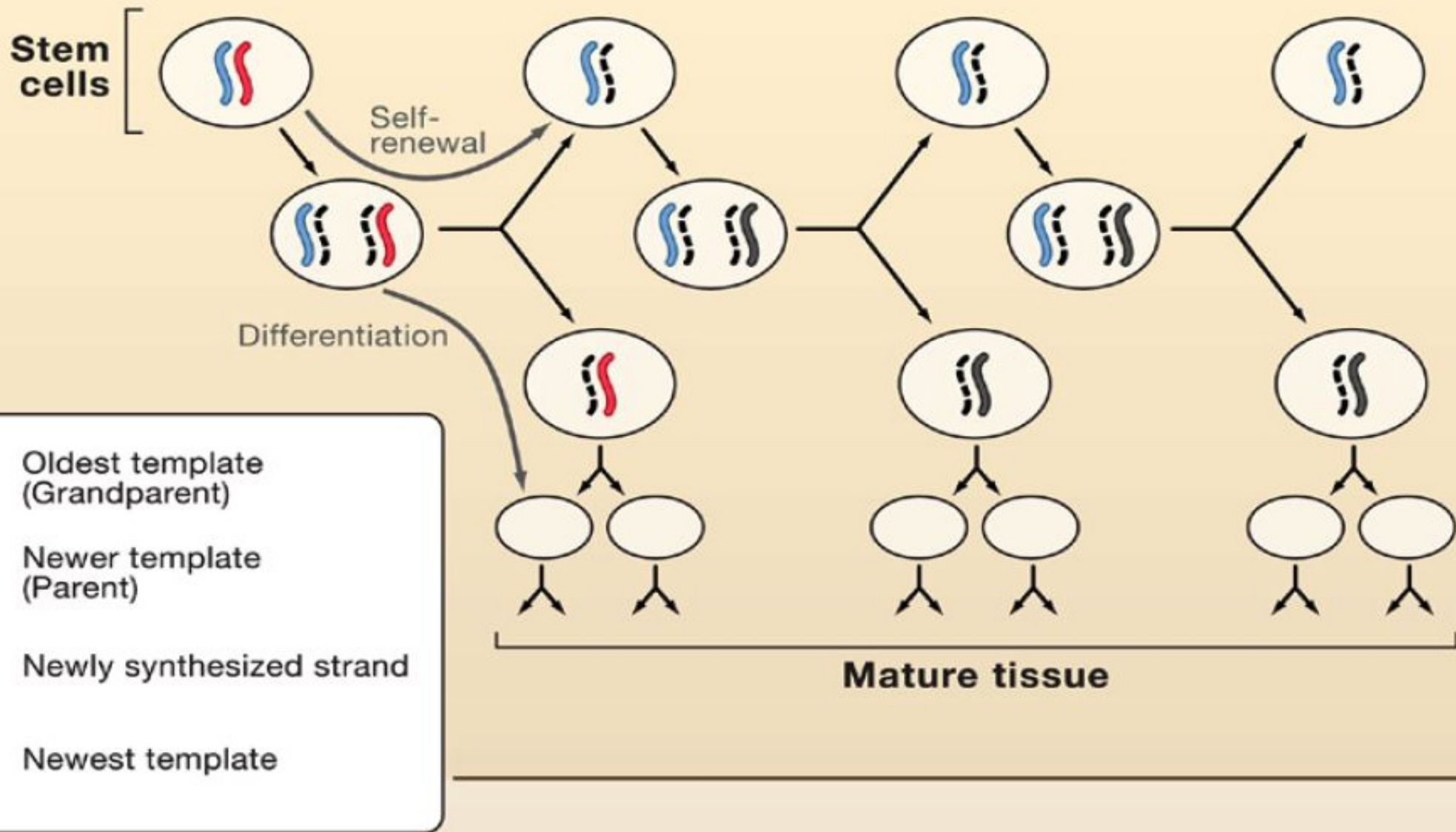


**Figure 2.** Evidence of Co-Segregation of DNA Template Strands during Muscle Progenitor Cell Division  
(B) Cell pairs were immunostained for CldU and IdU. Shown is a representative photograph of an immunostained pair of cells, in which both daughter cells were labeled with the second label, IdU (green), but only one daughter inherited the first label, CldU (red).

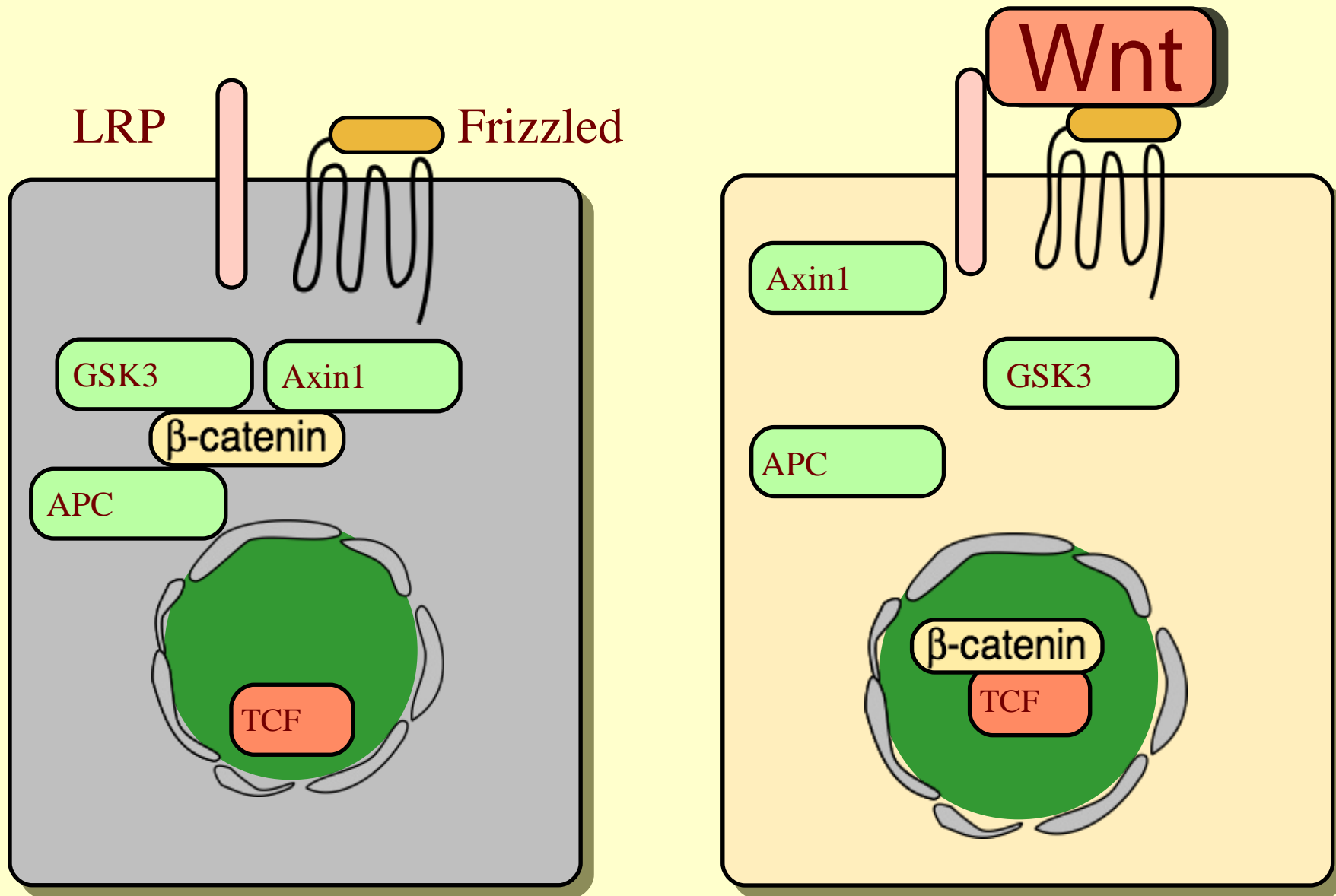




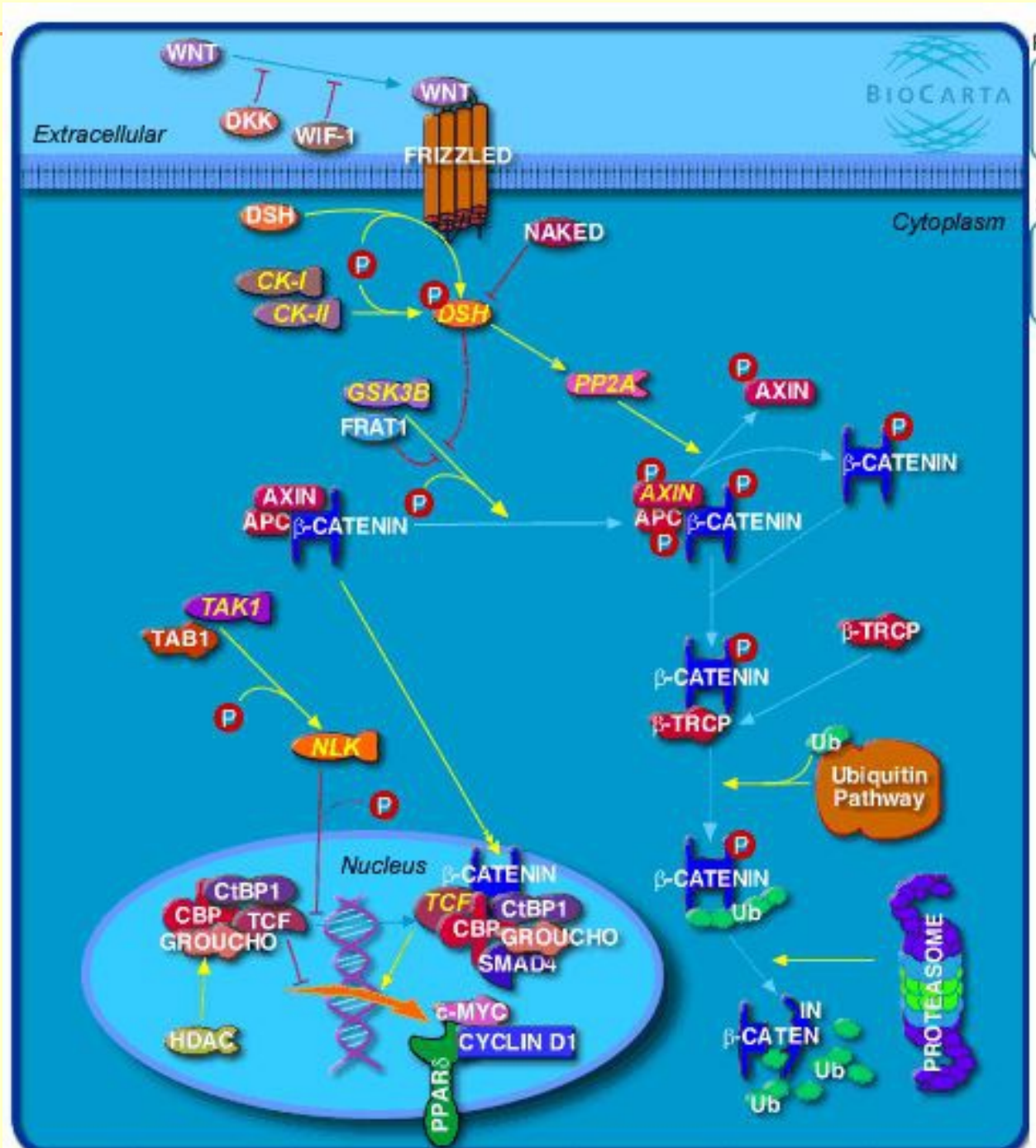
# Asymmetric Stem Cell Growth with Asymmetric Parental Strand Segregation



# Wnt signaling

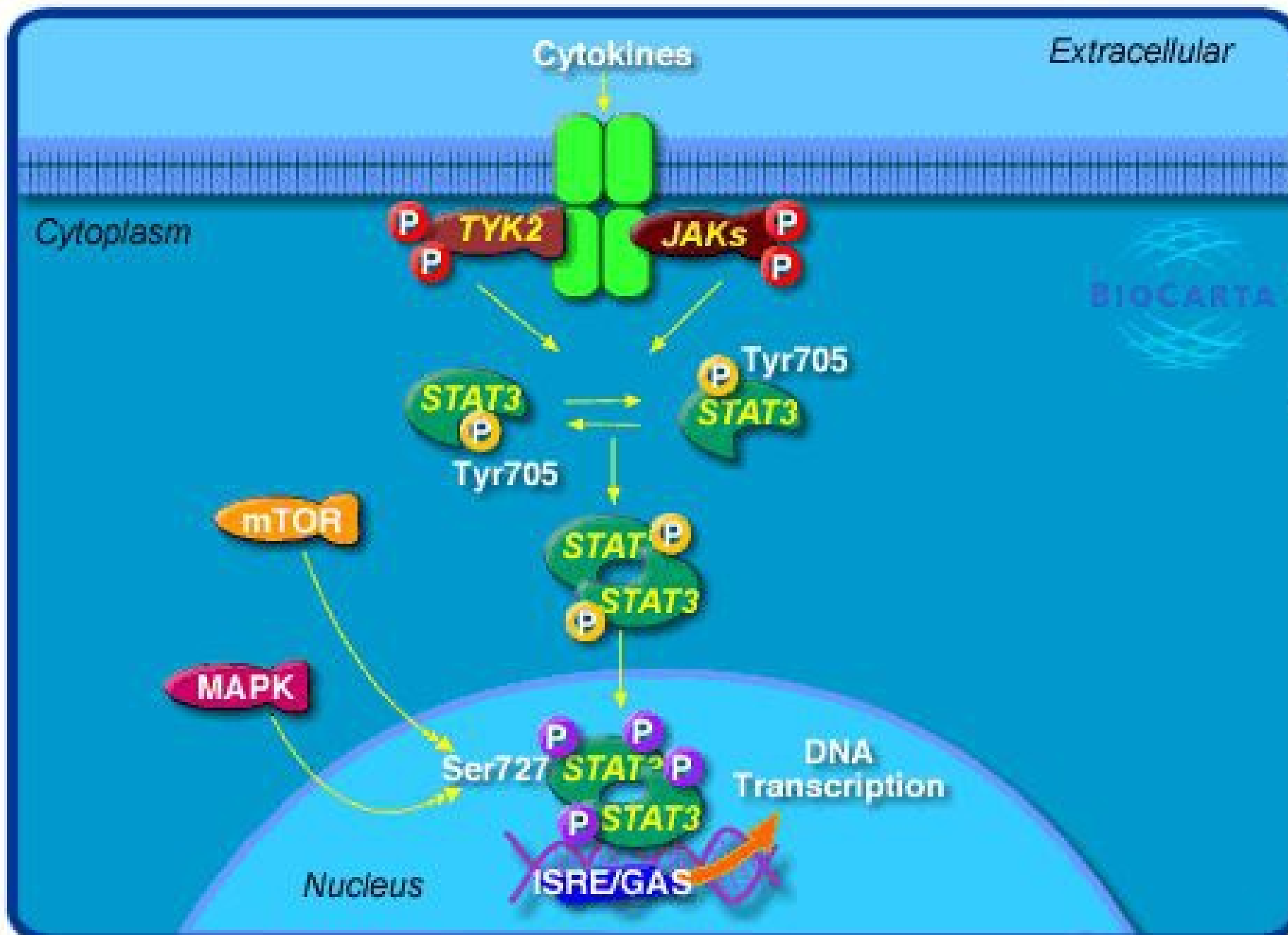


# Wnt Signaling Pathway

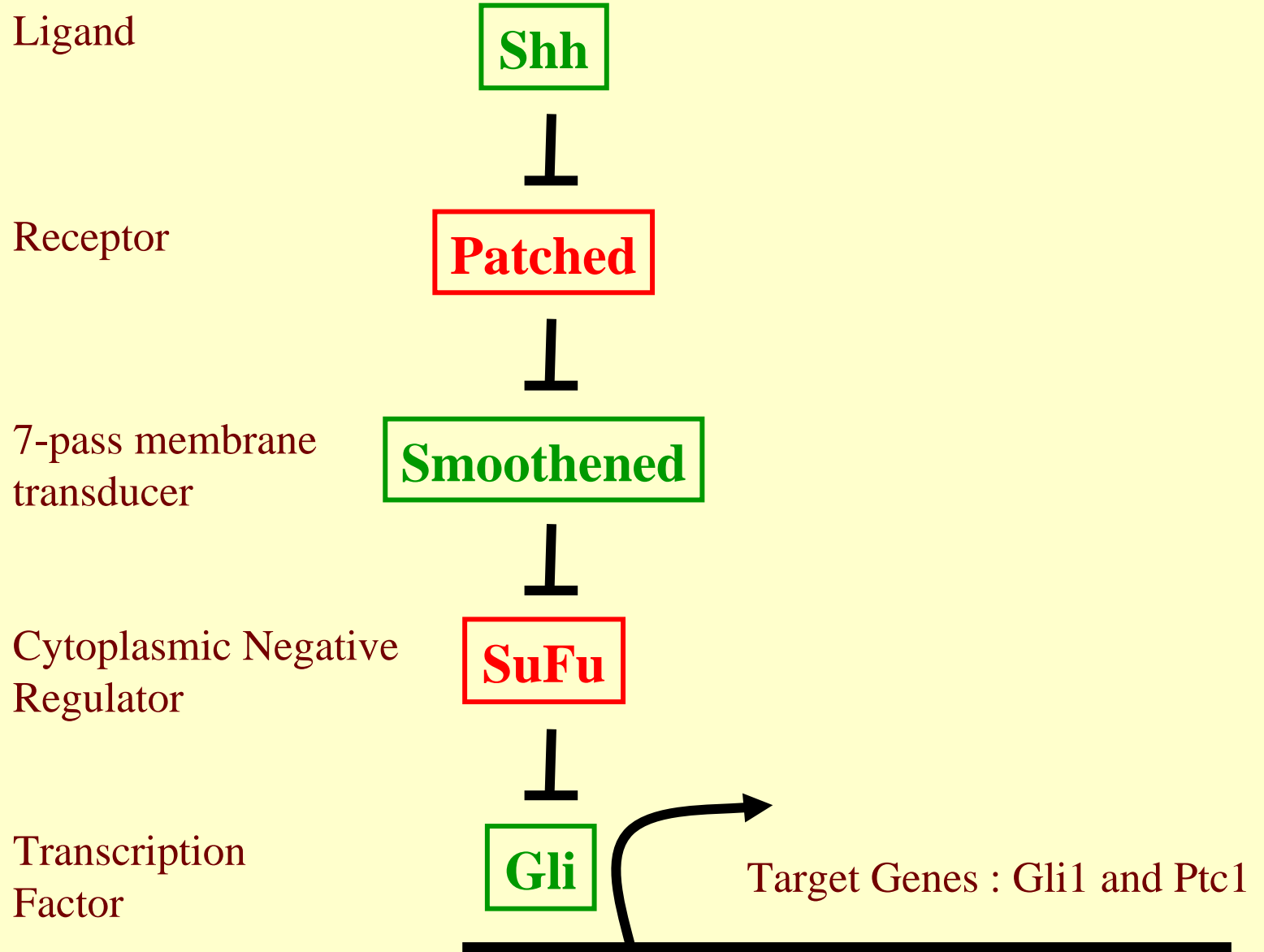


# Jak Stat Pathway

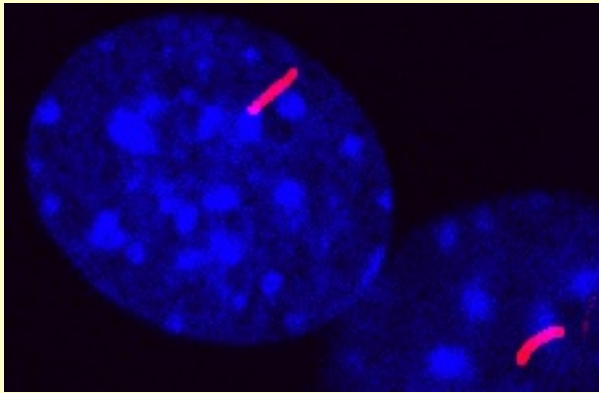
[http://www.biocarta.com/pathfiles/h\\_stat3Pathway.asp](http://www.biocarta.com/pathfiles/h_stat3Pathway.asp)



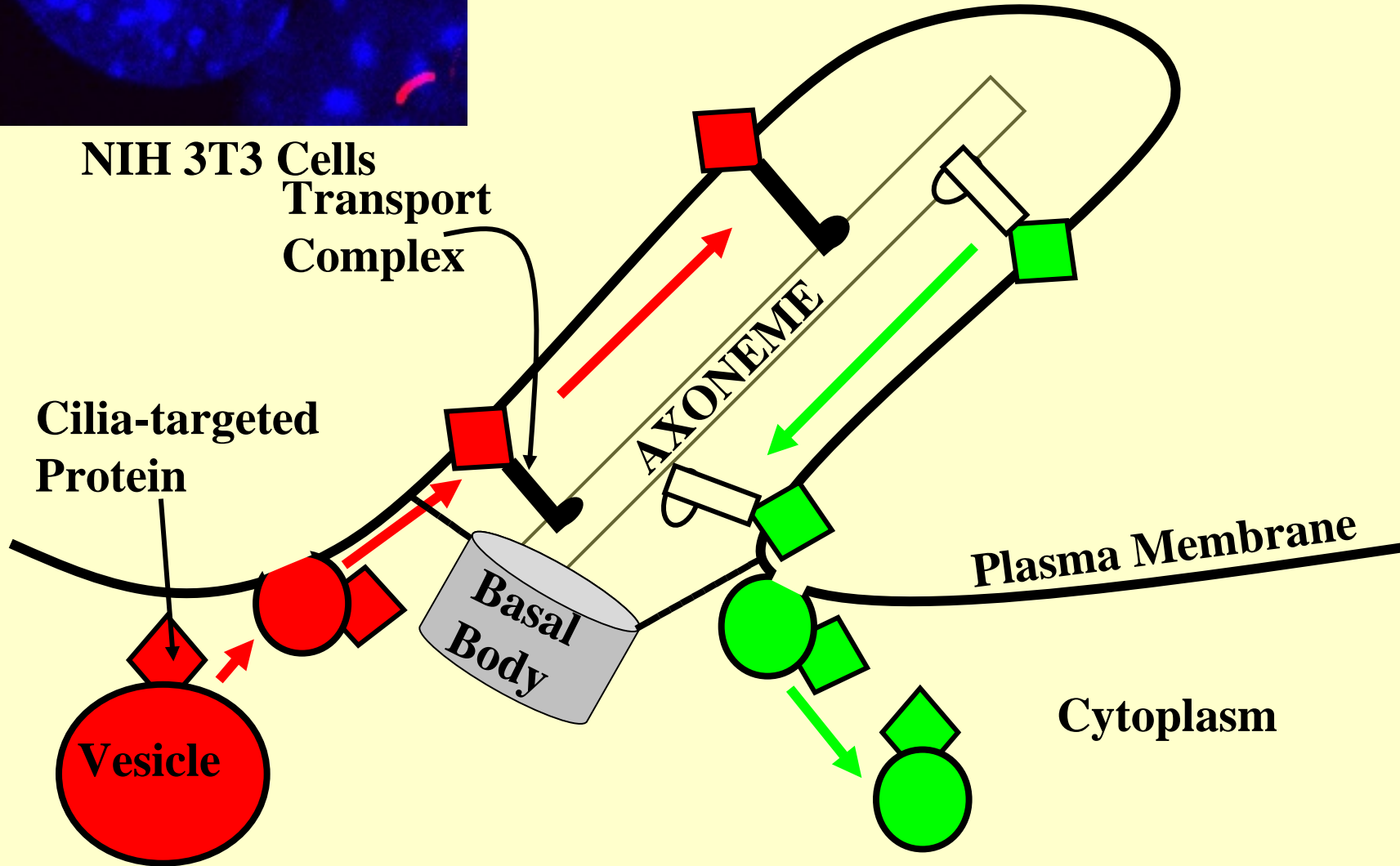
# The Hedgehog pathway



**Nucleus** **Cilium**

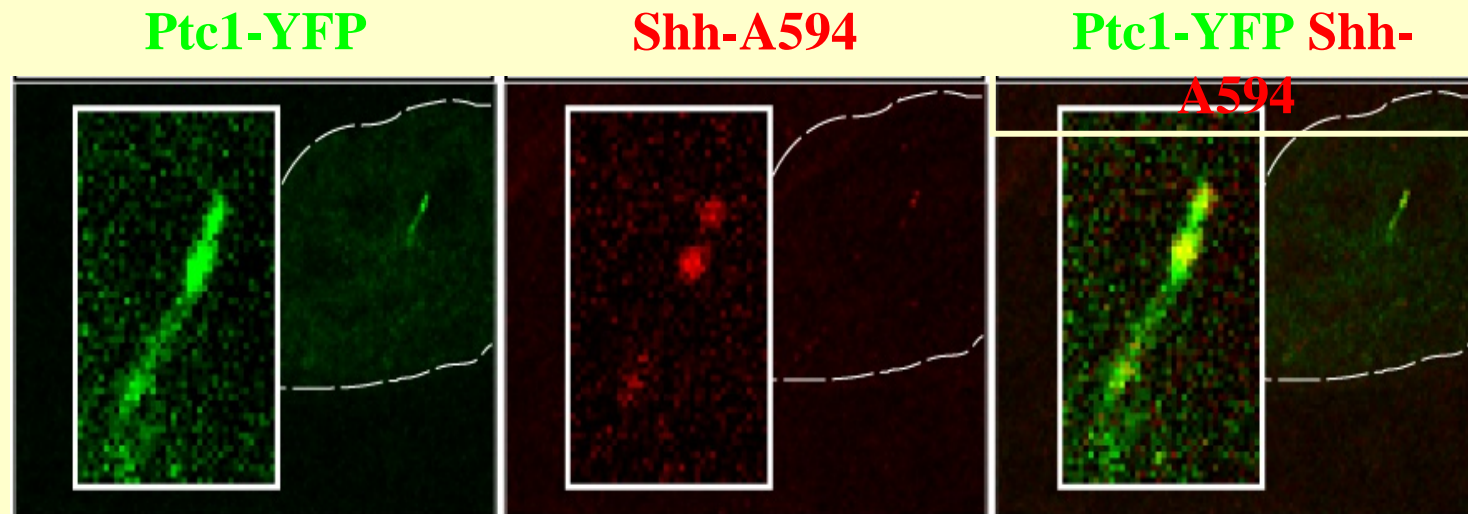
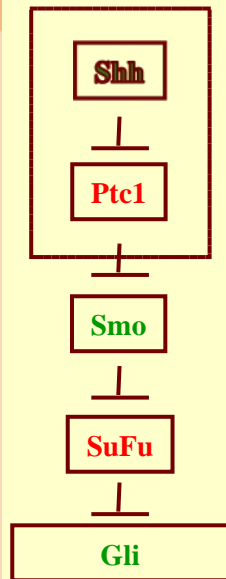


**NIH 3T3 Cells**  
**Transport**  
**Complex**

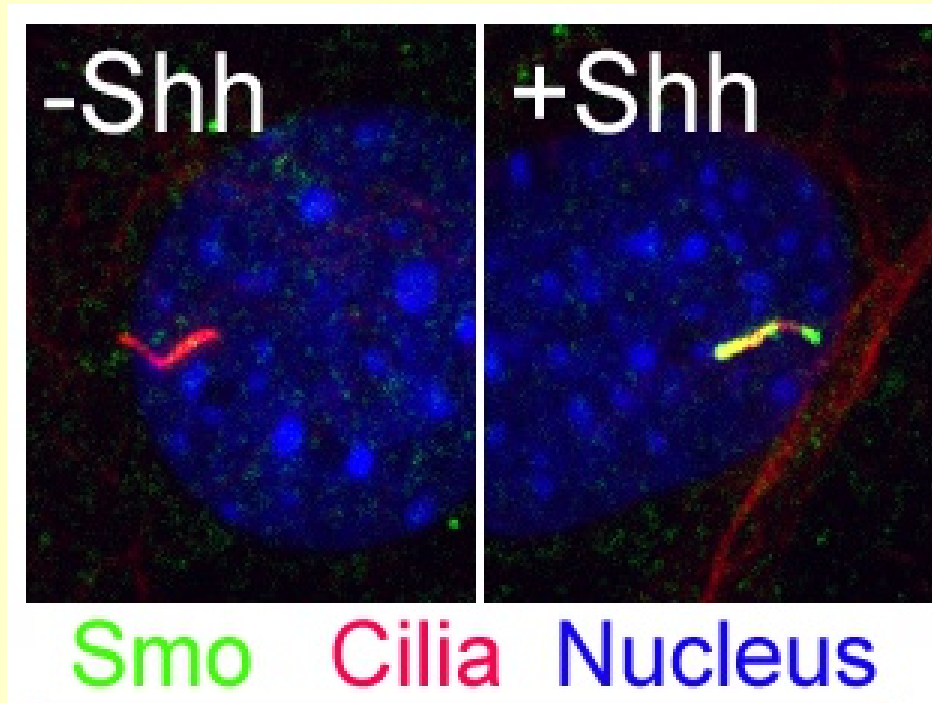
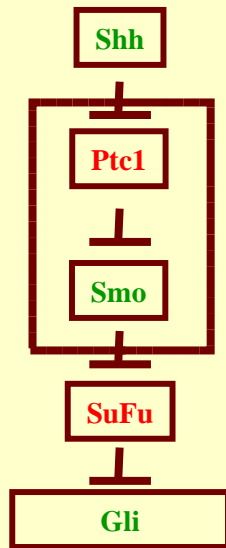


The primary cilium: A specialized compartment for signal transduction

# Cilia as sensors for Shh: Shh binds to its receptor Patched1 at primary cilia in live cells

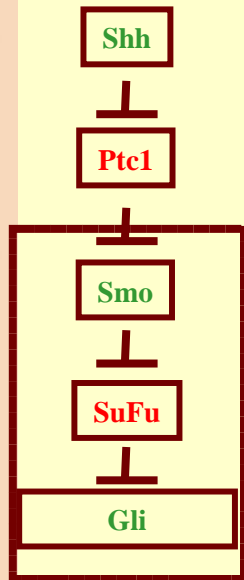


# Smo moves to cilia and when the Hedgehog pathway is activated

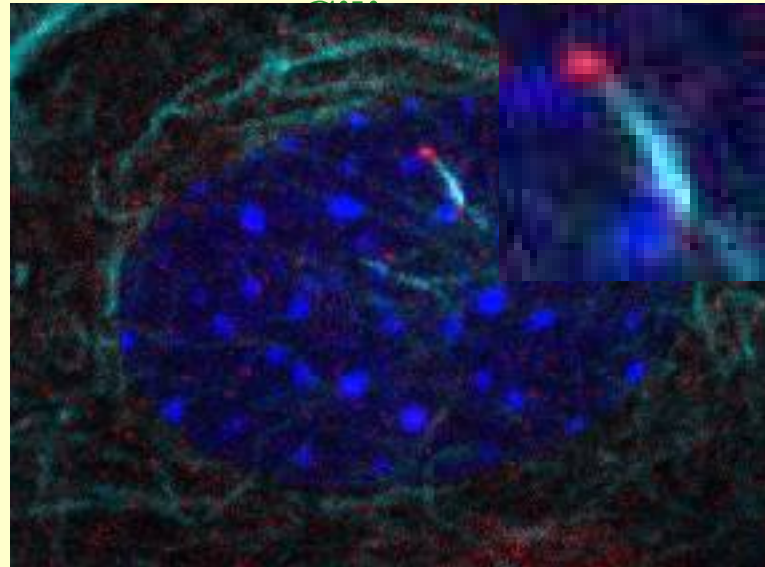




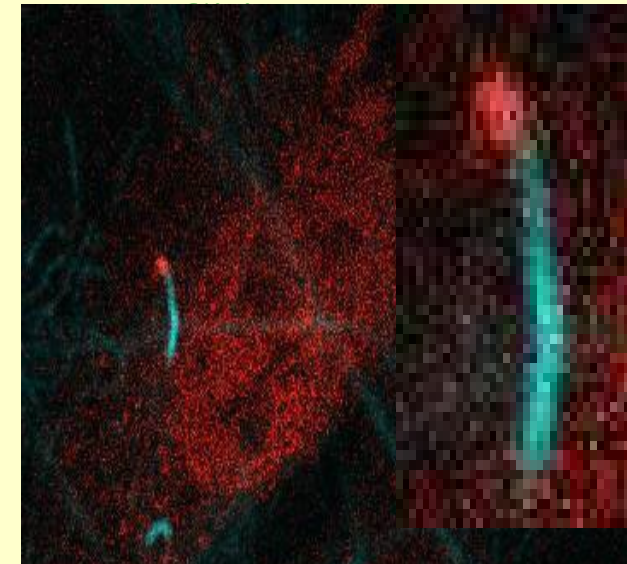
# Smo activates downstream signaling components in cilia



**SuFu**



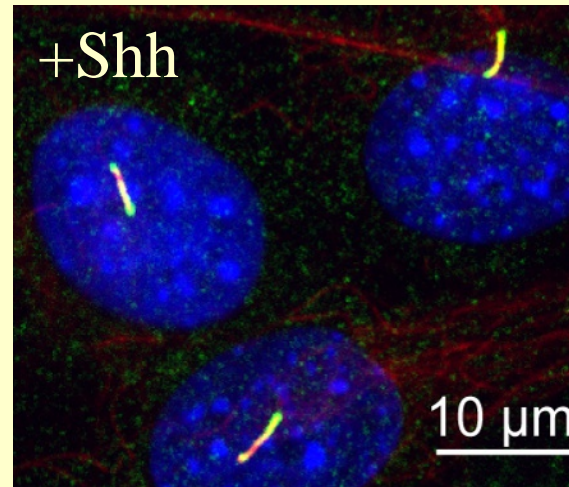
**Gli2-YFP**



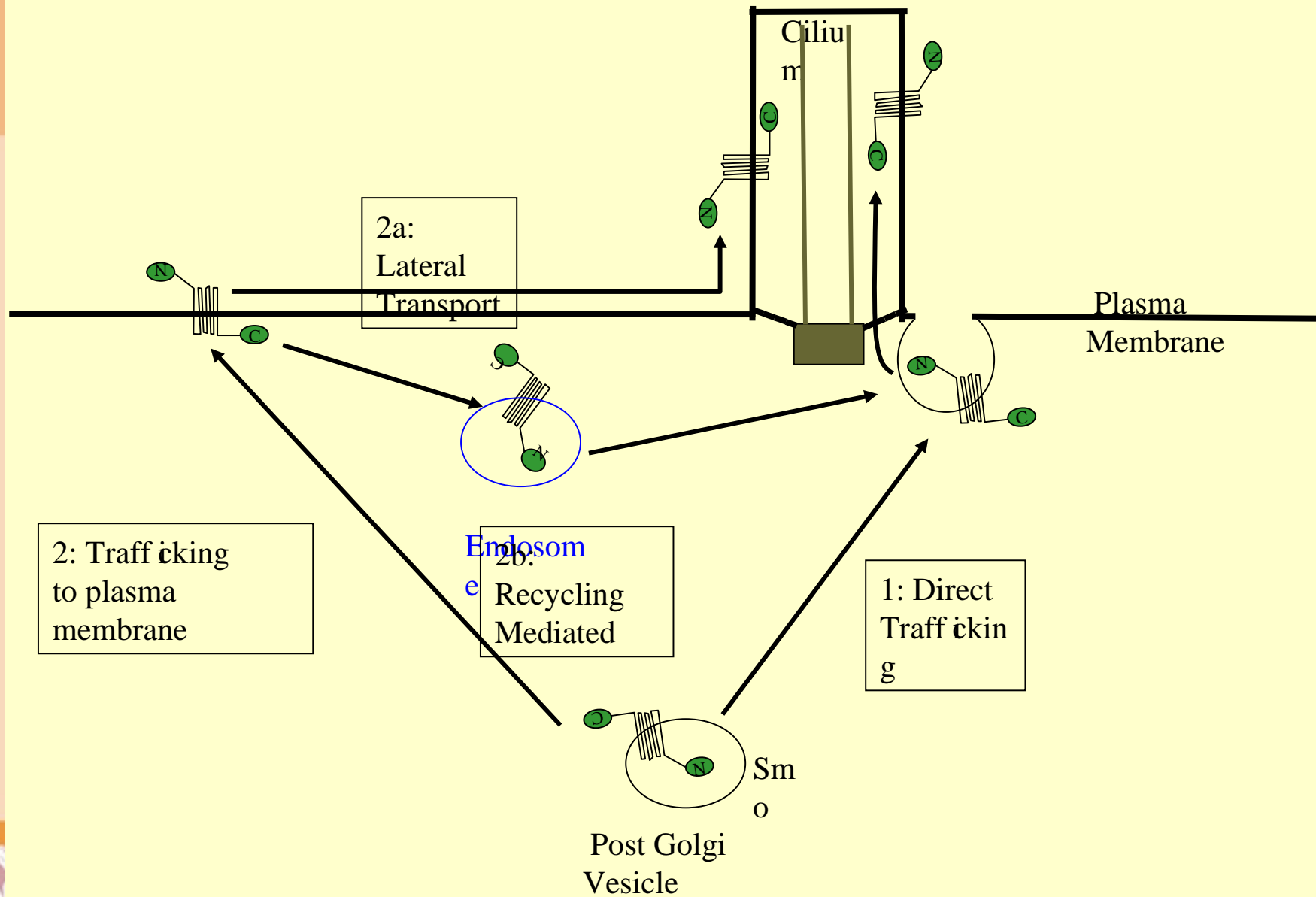
# Smo as a model for signal-regulated protein transport at primary cilia

---

**Smo** **Cilia** **Nucleus**

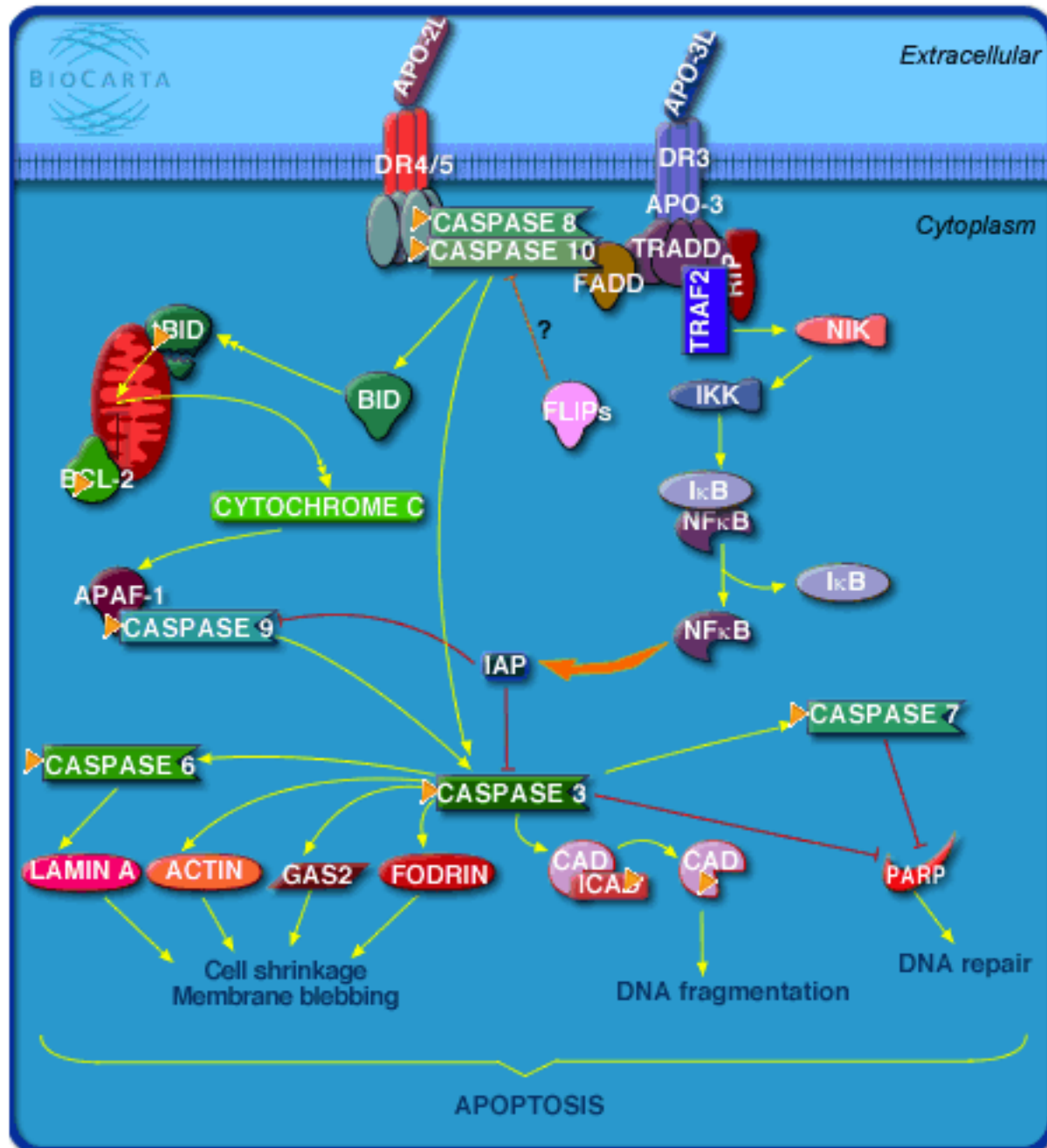


# Models for ciliary protein transport

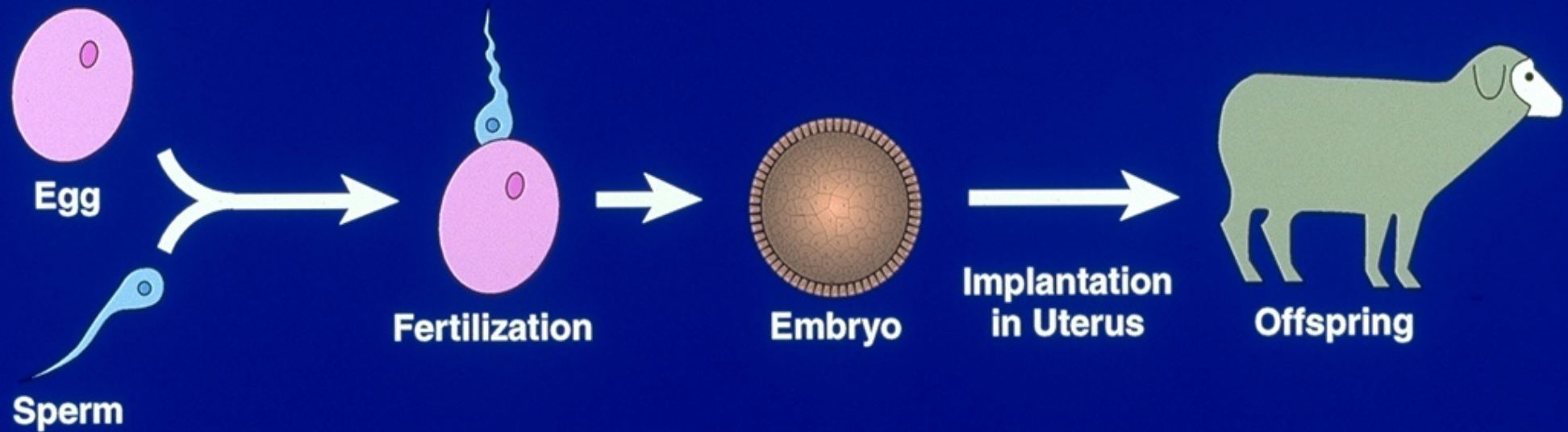


# Induction of Apoptosis via DR3 and DR4/5 Death Receptors

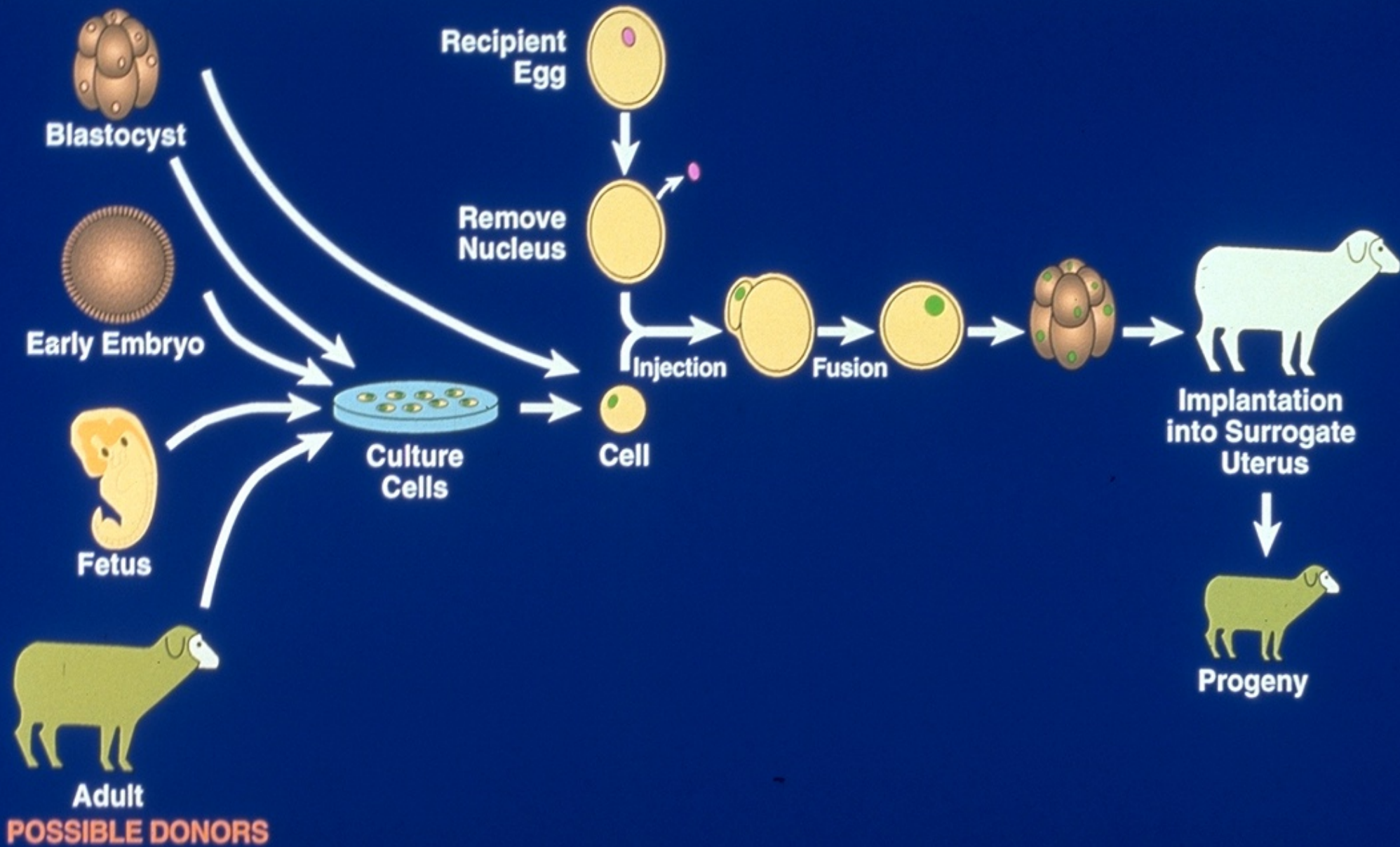
[http://www.biocarta.com/pathfiles/h\\_deathPathway.asp](http://www.biocarta.com/pathfiles/h_deathPathway.asp)

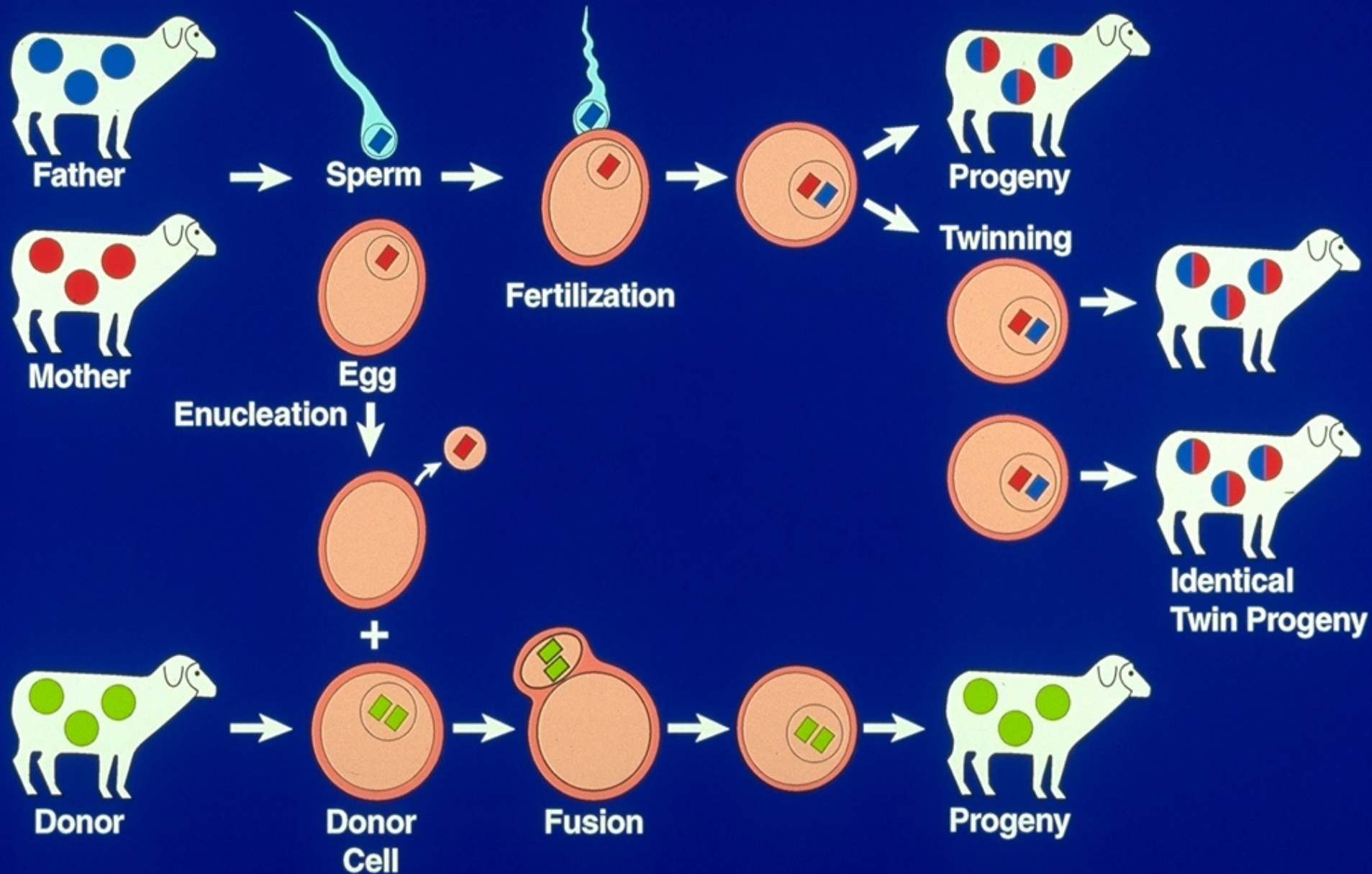


# Normal Reproduction

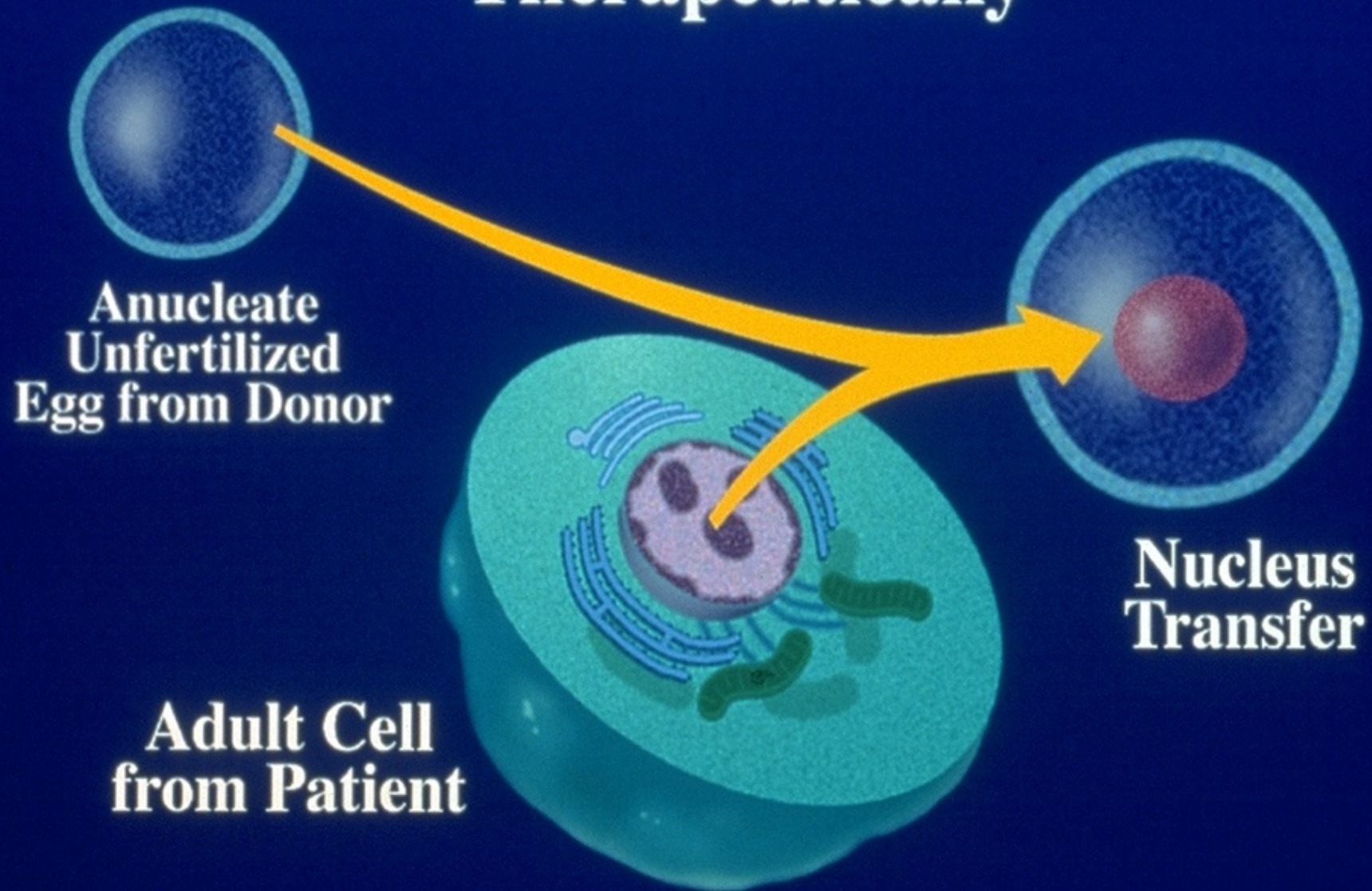


# Cloning Procedures





# How Cloning might be used Therapeutically



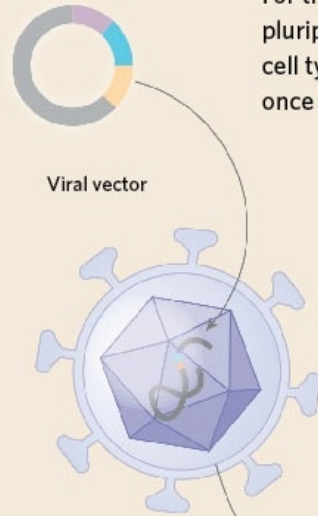


# Direct versus indirect Cell Reprogramming

<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>

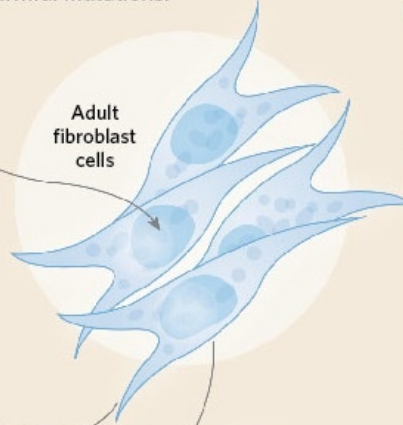
## CELLULAR REPROGRAMMING

For the better part of the past decade, researchers have been reprogramming adult cell types, either into induced pluripotent stem cells (iPSCs), which themselves can give rise to diverse cell types, or directly into other differentiated cell types through a process called direct reprogramming. Such approaches support the switching of diverse cell types once believed to be permanently locked in their differentiated form.



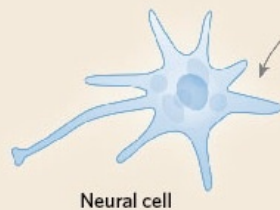
Viral vector

Traditionally, relevant transcription factors encoded by genetic material were carried by retro- or lentivirus vectors and integrated into the host cell genome. More recently, the use of nonintegrating vectors, RNA, or small molecules have been developed to minimize the chance of harmful mutations.

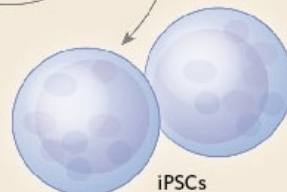


Adult fibroblast cells

Fibroblasts were the first and remain the most common type of cell to be reprogrammed, but other cells, such as lymphocytes, which can be isolated from blood, are also proving to be successful starting points for stem-cell generation.



Neural cell



iPSCs

Direct reprogramming into another adult cell type

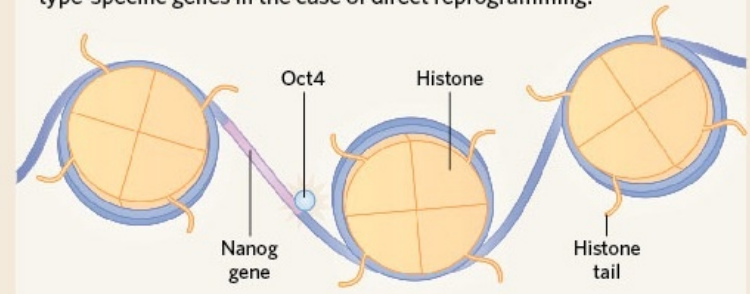
OR

Dedifferentiation into a pluripotent state

© LUCY READING-IKANDA

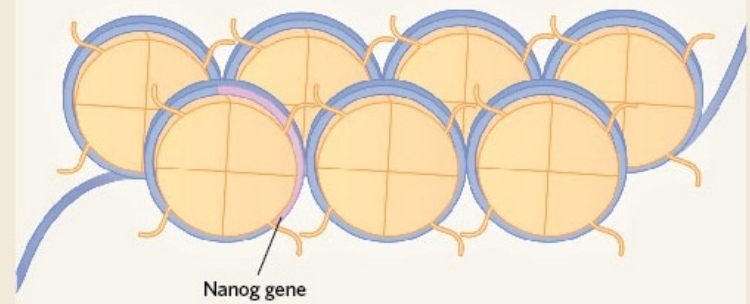
### OPEN CHROMATIN

Transfected transcription factors, such as Oct4, induce the expression of pluripotency-related genes, such as *Nanog*, or cell-type-specific genes in the case of direct reprogramming.



### CLOSED CHROMATIN

Sequences from pioneer factors, such as the myogenic factor MyoD, are also employed to increase reprogramming efficiency in the face of closed chromatin, which can inhibit access of the transfected transcription factors to their target genes.



<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>